

Intrinsic fluorescence in non-aromatic peptide structures is induced by collective vibrations, charge reorganisation and short hydrogen bonds, as shown in a new glutamine-related structure

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Abstract

Disentangling the origin of the optical activity of non-aromatic proteins is challenging due to their size and thus their high computational requisites. Here we show, in a much smaller model system, that the single amino acid glutamine undergoes a chemical transformation leading to an unreported glutamine-like structure which has a similar broad absorption spectrum reported previously for non-aromatic proteins. We further show computationally that the optical activity of the glutamine-like structure is directly coupled to short-hydrogen bonds, but also displays charge and vibrational fluctuations, the latter of which are also present in less optically active structures such as in L-glutamine. Since experimentally the glutamine-like structure is the brightest structure, we conclude that short-hydrogen bonds are the ones responsible for the large Stokes shift observed in optically active non-aromatic proteins.

38 Introduction

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40 Short peptides void of any aromatic residues have been shown to display an intrinsic
41 fluorescence in the visible range (1, 2). This has primarily been observed in fibrillar protein
42 structures linked to neurodegenerative diseases, such as Alzheimer's, Parkinson's and
43 Huntington's diseases (3–6). Furthermore, optical properties of double amino acid based
44 nanowires have also been reported, existing either of two non-aromatic or two aromatic
45 amino acids (2, 7, 8). We have shown that the fluorescence of non-aromatic short crystal
46 structures forming part of the amyloid-beta protein is enhanced by the presence of short
47 hydrogen bonds (SHB) between the termini of prefibrillar structures (5). The SHB permits
48 proton transfer leading to a double-well ground state potential which we have proposed
49 prevents a conical intersection in the excited state (5). It has been suggested that one of the
50 prerequisites for this fluorescence observed in either amyloid structures or short peptide
51 nanowires is related to hydrogen bonding or aromatic interlocks which, for the latter,
52 decreases the bandgaps down to the semiconductive regions (9).

53

54 Despite our previous suggestion that proton delocalisation is strongly coupled to this
55 intrinsic fluorescence, its direct role and more importantly, the role of other vibrational
56 modes on putative fluorescing states, has not been elucidated. We have thus searched for a
57 model system, such as a single amino acid-based structure, that displays similar optical
58 properties to amyloid fibrils and is permissive to more sophisticated computational
59 approaches. We have been inspired by the small peptide nanostructures that have been
60 pioneered by the Gazit laboratory (9) and by the fact that there are several
61 neurodegenerative diseases that have been connected with an increased level of glutamines
62 produced as part of a protein, as for example Huntingtin in Huntington's disease which
63 renders the protein more aggregation prone (10). It has been known that the amide group
64 in L-glutamine (L-glu) is highly labile and thus can rapidly hydrolyse. We show here that the
65 single amino acid L-glu upon heating in water can form a nanostructural material with
66 optical properties similar to the ones observed in other amyloid fibrils such as in fibrils of
67 amyloid-beta, alpha-synuclein or tau (4, 11, 12).

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Using X-ray diffraction (XRD), we show that L-glu dissolved in water and upon heating becomes cyclised forming a previously unreported structure which resembles L-pyroglutamine (which has been reported to be a component of amyloid-beta in the brain (13)), but involves a low-barrier hydrogen bonded anionic dimer with an ammonium counterion. We have termed the new structure, i.e. L-pyroglutamine complexed with an ammonium ion, L-pyro-amm. L-pyro-amm has a microcrystalline plate morphology as shown by scanning electron microscopy (SEM). The newly formed solid was further characterised using terahertz time-domain spectroscopy (THz-TDS), which provides information on the low-frequency modes in the crystal that control the proton transfer. Additionally, the experiments were interpreted using ground and excited state electronic structure calculations and molecular dynamics simulations. Ultimately, the combination of static structural information, atomic vibrational dynamics, and optical properties enable the origins of fluorescence in this particular structure to be elucidated, shedding light on the complementary processes in more complex systems.

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84 **Methods**

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86 *Experimental*

87 *Sample preparation of L-glutamine*

L-glutamine (L-glu) (#G3126, #G8540, Sigma-Aldrich, Gillingham, UK) and L-pyroglutamine (L-pyro) (#83160, Sigma-Aldrich) were dissolved in 18.2Ω MilliQ H₂O at a concentration of 0.3 M or 1 M. Aliquots were placed in a 65°C oven, since heating up proteins to 65°C increases the formation of amyloid structures as reported previously (14). Each aliquot was rotated to dissolve the powder once a day. Samples were either analysed in liquid form or dried on a glass or quartz cover slip (#043210.KG, Alfa Aesar, Lancashire, UK) either at room temperature (RT) or on a heat block set to 50°C.

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96 *Emission and excitation wavelength scans*

Emission and excitation spectra were taken on a Hitachi F-4500 FL spectrophotometer (Hitachi High-Technologies Corporation, Tokyo, Japan) at RT in a quartz cuvette. For measurements, the excitation slit resolution was 5 nm or 10 nm and the emission slit resolution was 20 nm. The PMT voltage was set at 950 V and the scan speed set at 240

nm/min. The excitation scan was measured between 250 – 400 nm and the emission filter set to the emission maxima of the sample stated in the figure legend, with a slit resolution of 20 nm. The emission scan was measured between 380 - 560 nm and the excitation filter set to the excitation maxima of the sample stated in the figure legend, using a slit resolution of 5 nm. Four measurements were taken for each sample which was repeated at least three times and the background (air or H₂O) was subtracted from the average.

Absorption measurements

Absorption measurements were taken on a UV-Vis-NIR Spectrophotometer, UV-3600 Plus (Shimadzu, Kyoto, Japan) and Cary 6000i (Agilent, Santa Clara, USA). 1 M or 0.3 M L-glu, L-pyro or L-pyro-amm solutions were measured in 10 mm QX cuvettes (Hellma Analytics, Müllheim, Germany) or dried on quartz coverslips. Measurements were taken between wavelengths 200 – 800 nm using 1 nm steps at a slow scan speed and a 1 nm resolution. The light source change wavelength was set at 393 nm and the grating change wavelength set at 750 nm. Samples were measured at least three times and the experiments repeated at least three times, measurements were then averaged and H₂O or coverslip only control was subtracted.

SEM (scanning electron microscopy)

SEM was performed using a FEI Magellan 400 HR-SEM at an acceleration voltage of 2 kV. L-pyro-amm samples were lyophilised by freezing in liquid nitrogen and freeze drying in a LyoQuest 85 (Telstar, Terrassa, Spain) and imaged on a glass coverslip.

X-ray diffraction

L-pyro-amm was dried on a glass coverslip in a 50°C oven and then at RT until crystals formed. Single crystal X-ray diffraction (SCXRD) measurements were performed at 180 K with a Bruker D8-QUEST PHOTON-100 diffractometer, which utilised a Cu K α radiation (λ = 1.54 Å), and an APEX-II CCD. Absorption corrections were made using SDABS, and data integration and reduction were performed with SAINT+. All non-hydrogen atoms were refined isotropically and anisotropically, followed by inclusion of the hydrogen atoms (determined using the excess electron density) and refinement isotropically.

Terahertz Time-Domain Spectroscopy

All THz-TDS spectra were acquired using a commercial Terapulse 4000 spectrometer (TeraView Ltd, Cambridge, UK). Samples were prepared for THz-TDS measurements by diluting the solid air dried L-pyro-amm with polyethylene (~ 10% w/w concentration) by gentle mixing using an agate mortar, followed by pressing into 2 mm thick, 13 mm diameter pellets using a hydraulic press. All THz-TDS spectra shown are a result of division of sample and blank datasets, with the blank dataset represented the THz-TDS response of a pellet of pure polyethylene.

Theoretical

DFT-THz Calculations

Calculations were performed using both the CRYSTAL17 (15) and Quantum Espresso (16) software packages. Geometry optimisations and vibrational analyses performed with the CRYSTAL17 code utilised the atom-centred split-valence triple-zeta 6-311g(2d,2p) basis set for all atom types. Based on a previous study related to ionic molecular crystals (17), the range-corrected WB97-X (18) functional was used. The vibrational analysis was performed within harmonic approximation, and infrared intensities were determined using the Berry Phase method. Energy convergence criteria were set to $\Delta E < 10^{-8}$ and 10^{-11} hartree for the geometry and frequency calculations, respectively.

Periodic TD-DFT Excited State Calculations

Simulations were performed using the fully periodic Quantum Espresso software package. The Becke-Lee-Yang-Parr (B3LYP) hybrid density functional was used with an energy cutoff of 40 Ry. The calculations of the excited state were performed within the framework of TDDFT using the Liouville-Lanczos formalism implemented in the freely available Quantum-Espresso package (19). In this approach, the optical spectra are obtained directly over the wide spectral range without taking into account the numerically complex calculations of the single excited states. We used plane wave basis set and the electron-ion interactions were taken into account via norm conserving Martins-Troullier pseudopotentials (20). To determine the ground state wave function, we used the gamma point of the Brillouin zone. All the periodic calculations employed the computationally demanding B3LYP (21) hybrid

functional, the kinetic energy cutoff of 40 Ry was used for the wave functions. The intrinsic band width for the spectra was set to 0.003 Ry (~0.0408 eV).

Periodic Structure Geometry Optimisation

The structures obtained from the experiments were first geometrically optimized at 0°K using the Broyden-Fletcher-Goldfarb-Shanno (BFGS) minimisation algorithm implemented in CP2K (22, 23) package. A convergence criterion for the wave function optimisation was used as 5×10^{-7} a.u. Applying the method of the Gaussian and plane wave, the wave function was expended in the Gaussian double-zeta valence polarised (DZVP) basis set. The cutoff for the electronic density was set to 300 Ry. We used the gradient correction to the local density approximation and the core electrons were treated via Goedecker-Teter-Hutter pseudopotentials (24). In all the calculations, we used the Becke-Lee-Yang-Parr (BLYP) (25) functional with the D3(0) Grimme (26) dispersion corrections for the van der Waals interactions.

Ab Initio Molecular Dynamics Simulations

Ab initio Molecular Dynamics simulations (AIMD) were performed using Quickstep algorithm implemented in CP2K. In these calculations, the propagation of the nuclei was taken into account within the framework of the Born-Oppenheimer approximation. The simulations were performed in the NVT ensemble and the temperature was controlled during the simulations by using the velocity-rescaling thermostat (27). We used the time step of 0.5 femtosecond to update the nuclear coordinates and velocities while the total length of the simulations for each system is 50 picoseconds.

Excited State Cluster Calculations

A set of excited state calculations were performed on glutamine clusters in order to understand the role of the environment on the optical properties. Specifically, the optical properties of L-pyro-amm were investigated using various isolated cluster models with the Gaussian09 software package. The clusters were extracted directly from the crystal structure and used in various combinations (dimers, trimers, tetramers) to perform time-dependent DFT (TD-DFT) calculations. A split-valence triple-zeta 6-311g(2d,2p) basis set was used for all atom types together with the hybrid B3LYP functional. Some benchmark

simulations, comparing the optical properties obtained from the periodic calculations using B3LYP to range corrected hybrid functionals like CAM-B3LYP, were also performed with these clusters.

We also performed a series of excited state optimisations on various model systems built from L-pyro-amm in order to examine the nature of the geometrical distortions that occur on the lowest electronic excited state. These calculations were also performed with the Gaussian09 software package. All clusters were surrounded with a continuum dielectric constant of 80, representing pure H₂O. The 6-311G(2d,2p) basis set was used for all atoms together with the range corrected hybrid functional CAM-B3LYP (28). The clusters were first optimised in the ground state after which they were optimised on the first electronic excited state.

Results and Discussion

It has long been known that poly-glutamine can form amyloid-like fibrillar structures *in vitro*. The more glutamine residues in the poly-glutamine polymer, the faster the aggregation propensity of the polypeptide chain. This led us to investigate whether L-glu on its own under conditions which normally promote fibril formation, such as an increase in temperature (14), was able to form structures with similar optical properties, as recently observed for amyloid fibrils (5, 29, 30).

We first investigated the structure of L-pyro-amm, which formed after incubation of L-glu for 8 days at 65°C, using SEM and observed crystal structures shown in Fig.1a. However, in order to investigate whether L-glu had indeed changed its crystal structure arrangement we performed XRD analysis of the resulting material. In Fig.1b we show the crystal structure of the heated L-glu structure, which we termed L-pyro-amm, and the published crystal structures of L-glu and L-pyroglutamine (L-pyro) in Fig. 1c and d. Note, the L-pyro structure was analysed as it displayed structural similarities to the newly formed L-pyro-amm. Figures were obtained from geometry optimisations of the nuclear positions of the atoms using experimental densities. L-pyro-amm consists of 8 pyroglutamine groups and 4 ammonium ions (144 atoms) complexed within the crystal (see Fig. 1b). In contrast, as shown in Fig. 1c,

L-glu consists of 4 glutamine molecules (80 atoms) in the unit cell which form hydrogen bonds involving the termini and side chain. Furthermore, as shown in Fig. 1d, L-pyro consists of 12 pyroglutamine molecules (192 atoms) in the unit cell forming hydrogen bonds involving the NH and COOH groups.

L-pyro-amm has a rather unique hydrogen bond network structure since four of the pyroglutamine molecules are deprotonated and hence have a nominal negative charge, while the other four molecules are neutral. One of the important implications of this difference is that L-pyro-amm contains a very strong hydrogen bond. The red circled region in Fig. 1a corresponds to a short hydrogen bond (SHB) with a length of 2.45 Å, while those in L-glu and L-pyro range between ~2.55-2.85 Å.

The structural change was further confirmed using THz-TDS measurements, as this technique is strongly dependent on the bulk packing arrangement (as well as on the internal covalent structure) of the molecules (31). The THz-TDS spectrum of the resulting solid, as well as the solid-state DFT predicted spectrum based on the single crystal XRD (SCXRD)-determined structure, is shown in Fig. 1e (full spectral assignment available in Suppl. Fig. 1). The agreement between the experimental and theoretical spectra further supports that full conversion of the sample occurs and thus enables additional investigations into the structural and electronic properties of the material.

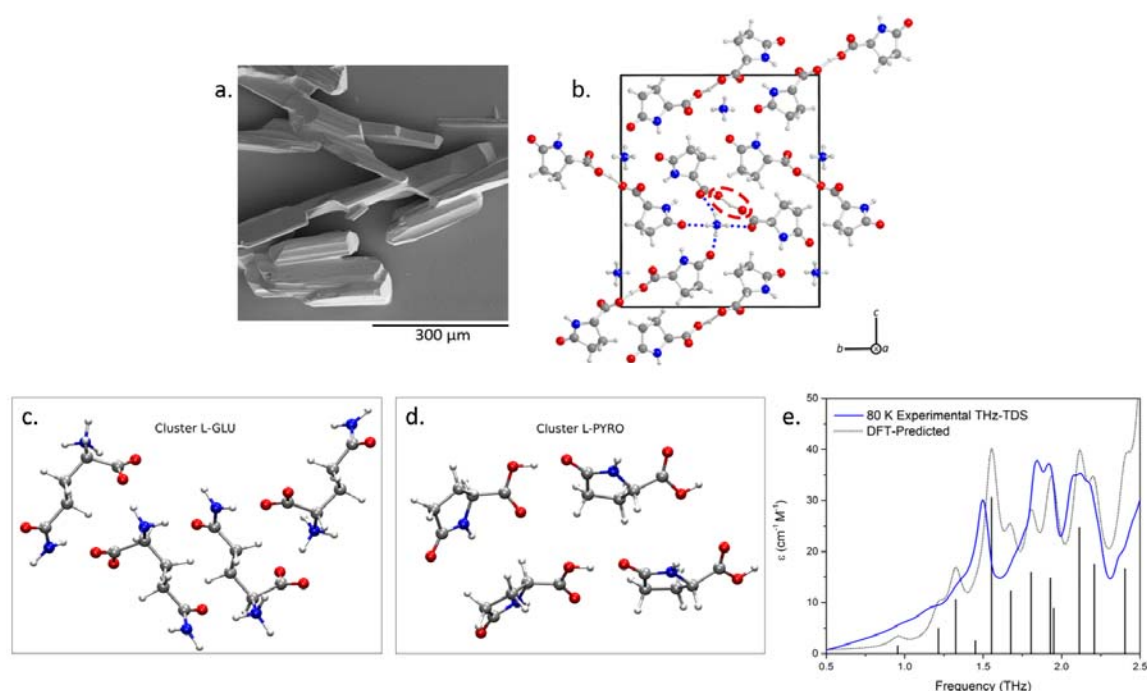


Figure 1. L-glu forms L-pyro-amm upon heating.

(a) SEM image of crystals of L-pyro-amm dried. (b) XRD analysis of heated L-glu sample show the newly formed structure, L-pyro-amm. Geometry optimisations show that 8 pyroglutamine groups and four ammonium ions (144 atoms) are complexed in the crystal and a SHB of 2.45 Å (within red dashed lines) is present near the ammonium ion (linked by bonds highlighted by blue dotted lines) (white-hydrogen, red-oxygen, blue-nitrogen, grey-carbon). (c) Clusters of L-glutamine and (d) L-pyroglutamine. (e) Experimental (blue line) and theoretical (grey line) THz-TDS of the L-pyro-amm sample are in agreement and confirm the presence of the new L-pyro-amm structure.

We first investigated whether there were any differences in the optical properties associated with the three crystal structures. Comparing the absorption of L-glu, L-pyro and L-pyro-amm in water, we show that only L-pyro-amm has a significantly red-shifted absorption which lies in the 275-320 nm range, whereas both L-glu and L-pyro primarily absorb in the deep UV (<250 nm) (see Fig. 2a).

We next compared the experimental absorption spectra of L-glu, L-pyro and L-pyro-amm with the ones obtained from time dependent density functional theory (TDDFT). We highlight here, that the small size of the systems permitted us to determine the spectra

using a hybrid functional, thereby not only advancing the quality of our theoretical predictions from previous studies (5, 29, 30) but also coupling the optical properties directly to different vibrational modes.

Fig. 2b illustrates the absorption spectra obtained for the TDDFT calculations on the 3 periodic systems in the ground state (i.e. at 0 K). Panel b.i) shows the relative oscillator strength as a function of the frequency while panel b.ii) illustrates the second derivative of the oscillator strength permitting the positions of the maxima in the spectra to be more easily identified. The spectra reveal some striking differences between the different systems. Interestingly, we observe that L-pyro is essentially dark throughout the frequency range up to ~ 6 eV. On the other hand, L-pyro-amm shows the presence of more structure in the spectrum. Specifically, it is the only system for which the spectrum features a low energy excitation at 226 nm (5.5 eV) and subsequently other peaks slightly above 220 nm (5.625 eV) and 216 nm (5.75 eV). While L-glu exhibits a peak at 222 nm (5.58 eV), it is dark up to 206 nm (~ 6 eV). These calculations were performed with the B3LYP functional. Importantly, we have found that upon using more accurate functionals such as CAM-B3LYP (28) and WB97-X (18) L-pyro-amm remains the most optically active.

We have previously shown that thermal fluctuations and in particular nuclear vibrations, such as proton transfer, have a large impact on the absorption spectra of peptide structures compared to absorption spectra at 0 K (5, 32–34). In Fig. 2c we show that, compared to the 0 K spectra in Fig. 2b, thermal fluctuations cause a large red shift to around 3.4 eV (365 nm) for L-pyro-amm, close to what is observed experimentally. These spectra were computed averaging over 25 frames sampled from the molecular dynamics simulations. Interestingly, no such effect is observed for L-glu which remains weakly absorbing up to more than 5 eV (247 nm) as seen at 0 K.

Similar to our previous studies on the intrinsic fluorescence of amyloid-beta fibrils, absorption is significantly increased in structures containing SHB, as neither L-glu, nor L-pyro display a significantly red shifted absorption. In order to understand better the physical origin of the low energy excitation at 226 nm (~ 5.5 eV) in L-pyro-amm, we computed the

electron response density at this frequency. This is illustrated in Fig. 2d, where we observe that most of the electron response involves regions around the pyroglutamine rings as well as regions near the SHB (see dashed circle in Fig. 2d). The optical response thus involves a collective charge reorganisation involving several parts of the molecular crystal.

We next investigated whether the above structures also display fluorescence excitation and emission properties as has been observed for amyloid-like structures reported previously (5, 29, 30). Fig. 2e shows the excitation scan from 250-400 nm (solid lines) with the emission set at 430 nm of L-glu in water at day 0 to 8 after incubation at 65°C. Interestingly, we observe an excitation peak at around 360 nm which is similar to what we have measured previously for amyloid proteins (5). The corresponding emission scan (dashed lines) with the excitation set at 360 nm and emission from 380-560 nm showed an emission peak around 430 nm, again lying in the same visible range as for amyloid fibrils. When the L-pyro-amm solution was dried the excitation and emission peaks were slightly blue shifted (Suppl. Fig. 2a) which may be due to a change in the molecular environment in the dried state. Importantly, we do not see any fluorescence in L-glu (without heating, i.e. at day 0 Fig. 2e.). To determine the importance of the ammonium ion experimentally, L-pyro has been incubated in water and heated at 65°C for 8 days, and only a very weak fluorescence has been detected (Suppl. Fig. 2b).

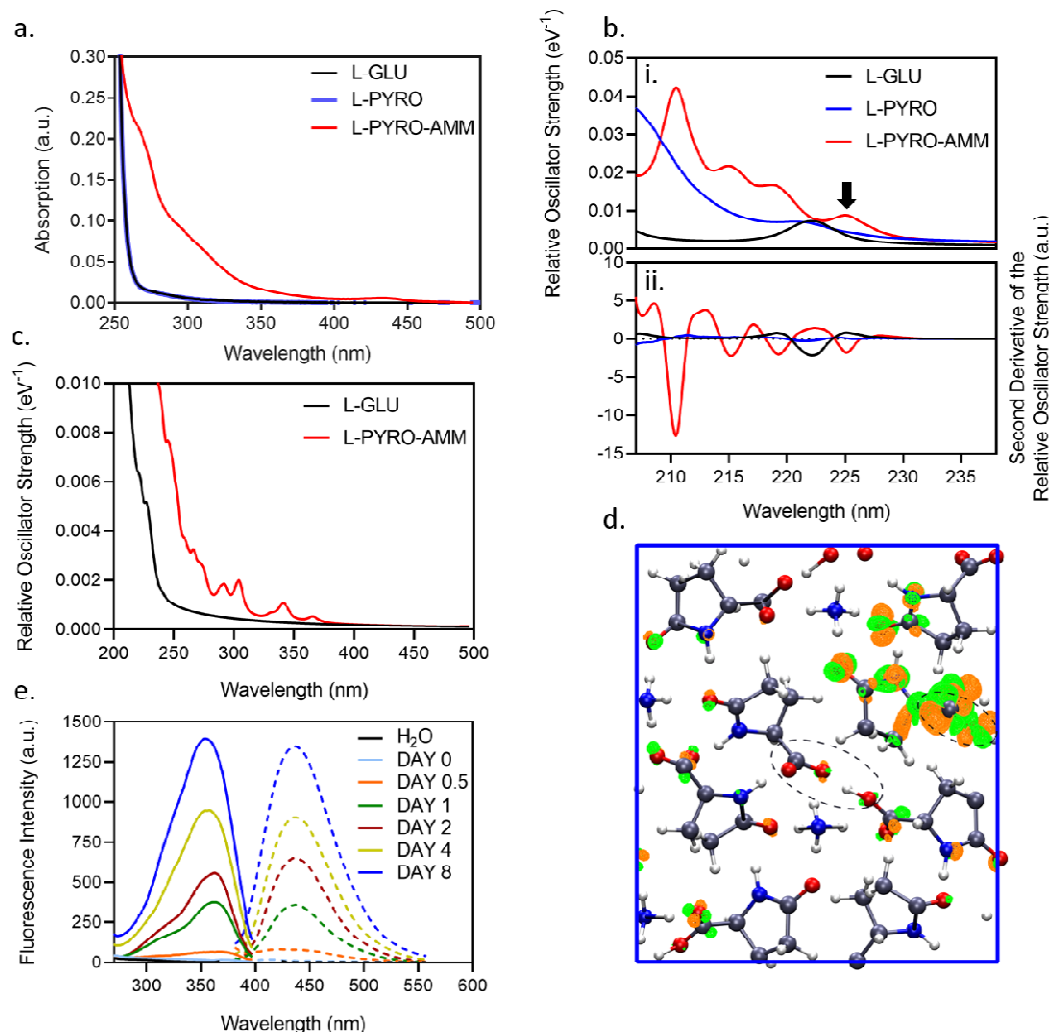


Figure 2. Optical properties of L-pyro-amm are distinct from L-glu and L-pyro

(a) Absorption spectra of 0.3 M L-glu (black), L-pyro (blue) and L-pyro-amm (red) (L-glu incubated for 8 days at 65°C) in water taken between 200 – 500 nm shows primarily features of L-pyro-amm. (b) Absorption spectra of L-glu, L-pyro and L-pyro-amm obtained from periodic density functional theory calculations with the B3LYP functional. L-pyro-amm features the lowest lying excited states which are characterised by the largest oscillator strengths. (c) Absorption spectra for L-glu and L-pyro-amm obtained from periodic simulations at room temperature. The spectra were computed by averaging over 25 frames randomly sampled from the *ab initio* molecular dynamics simulations. (d) The excited state electron density computed for L-pyro-amm from the optimised structure computed at the first peak (arrow in panel b). The lowest excited state density shows a response from various parts of the crystal structure including the pyroglutamic acid ring and the SHB region (see

dashed circle). The orange and green surfaces correspond to regions involving a decrease and increase in electron density respectively, shown at an iso-value of 1×10^{-5} . (e) 1 M L-glu was incubated at 65°C and the excitation and emission spectra were measured over time. Excitation spectra were measured between 250-400 nm with emission set at 420 nm and emission spectra were measured between 380-560 nm with the excitation set at 360 nm.

It is not possible to directly pinpoint the exact mechanisms and origins of the difference in the optical properties between the three systems from the ground state calculations at 0 K. As alluded to earlier, one of the factors that distinguishes L-pyro-amm from the other systems is the presence of the SHB (highlighted by red circles in Fig. 1a) and the presence of the ammonium ion. In order to characterise the behaviour of the SHB, we conducted *ab initio* molecular dynamics simulations of the three systems at 300 K and examined the proton transfer coordinates defined as the difference in distance between the proton (H) and the two oxygen atoms (O1 and O2) that sandwich it and is commonly referred to as the proton transfer coordinate ($d_{O1-H} - d_{O2-H}$) as shown in Fig. 4a for different types of hydrogen bonds in the crystals. It is clear that the SHB in L-pyro-amm is characterised by a double-well potential. The barrier associated with this proton transfer is on the order of thermal energy, indicating that zero-point energy (ZPE) would make the proton transfer barrierless (35). An examination of similar proton transfer coordinates for hydrogen bonds in L-glu and L-pyro show that they are characterised by only single-well potentials.

The nature of the optical properties is sensitive to the environment in which the glutamine molecules reside. It has previously been reported that charged amino acids already display an absorption in the range of 250-350 nm that is significantly red shifted (36, 37). The origins of the low energy absorption were attributed to charge transfer excitations. The simulations of these systems were performed in the gas phase, rather than considering the protein environment such as shown for L-pyro-amm in Fig 2d. In comparison to the results presented in Fig. 2d, data presented in Fig. 3, show that the origins of the electronic transitions equally arise from a charge transfer (CT) between the highest occupied molecular orbital (HOMO) on the anionic dimer, and the lowest unoccupied molecular orbital (LUMO) centred on the ammonium cation when performed in the gas phase. Interestingly, the correct transition energy is only predicted when the ammonium cation is

spatially near the centre of the dimer, which corresponds to the delocalisation of the negative charge and the SHB. Two generalised geometries, with the ammonium cation near the SHB (as seen in Fig. 3a) and away from the SHB (Fig. 3b), with the corresponding HOMO and LUMO orbitals are shown. The results predict a transition of 303.50 nm for dimer a. and 668.66 nm for dimer b., with dimer a. most closely resembling the chemical environment present within the crystalline material.

The results show that charge transfer is capable to lead to absorption in the far UV when investigated in the gas phase, i.e. neglecting the direct protein environment. However, including the protein environment in the molecular crystal (Fig. 2d) results instead in the excitation being a charge reorganisation involving several different molecular groups of the crystal. Indeed, by shuffling the protons along the SHBs in the ground-state, we observe an electronic response involving the entire structural units of L-pyro-amm including both the hydrogen bonded regions and the pyroglutamic acid rings when the protein environment is accounted for (Suppl. Fig.3b i-iv).

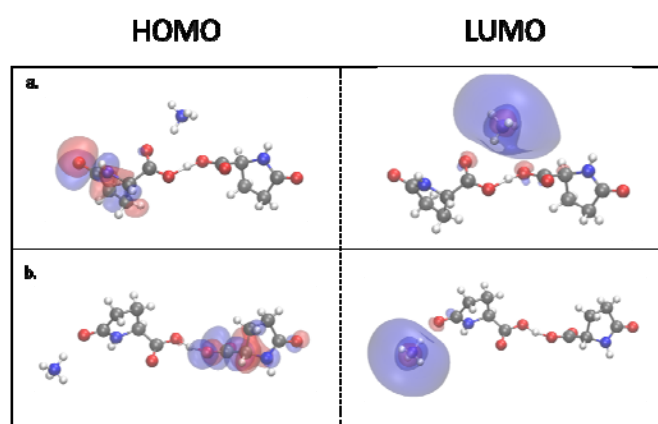


Figure 3. Comparison of the HOMO and LUMO orbitals on two L-Pyro-Amm models.

L-pyro-Amm structures are presented where the ammonium cation is located directly near the hydrogen bond (a), and where the ammonium cation is located away from the hydrogen bond (b). While both models predicted charge-transfer HOMO-LUMO states, only in the case of (a) is the transition predicted to be in the vicinity of the experimentally observed peak, 303.50 nm compared to 668.66 nm for (b).

Up to this point, we have shown that the vibrations of protons along SHBs are an important part of the structural fluctuations in the ground state structure of L-pyro-amm. These calculations however, do not say anything about the nuclear relaxation upon photo-excitation which is most relevant for fluorescence. In a final set of theoretical studies, to ascertain the role of the proton transfer in the ground state, as well as to elucidate the presence of other types of vibrational modes, we performed a series of geometry optimisations on the first excited state of a cluster carved out from L-glu, L-pyro and L-pyro-amm crystals and surrounded by a continuum dielectric constant of 80.

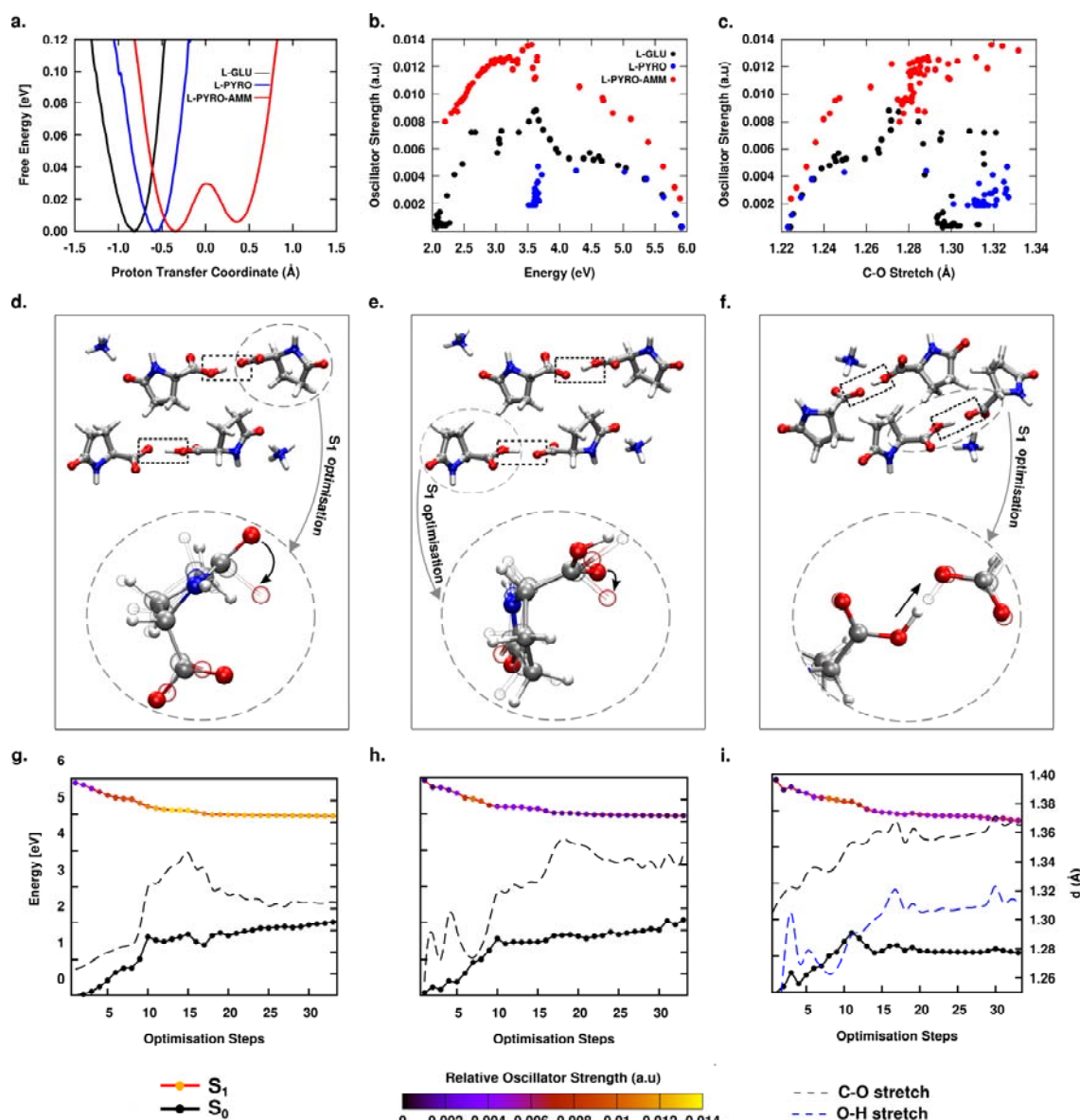


Figure 4. Collective vibrations affect the emission in L-pyro-amm

a) Shows the proton transfer free energy profiles along several hydrogen bonds in L-glu, L-pyro and L-pyro-amm obtained from ground-state ab initio molecular dynamics simulations. Note, the SHB in L-pyro-amm is characterised by a double-well potential. b) Scatter plot of the oscillator strengths versus the emission energy (defined as the difference between the first excited state and the ground-state) during the excited optimisations for L-glu, L-pyro and L-pyro-amm. c) Scatter plot of the oscillator strengths versus the distance of the carbonyl bond (C=O) that extends during the excited state optimisation in L-glu, L-pyro and L-pyro-amm. d)-f) Snapshots of three different systems of L-pyro-amm that were optimised on the excited state. In clusters shown in d) and e) the initial position of the proton along the SHB has been changed, and in f) the ammonium ion has been placed closer to the SHB region. The bottom panel of d), e) and f) show zoomed in plots of the main regions of the cluster (circled in the top panels) that undergo significant changes. g)-i) Ground and excited state energies are plotted as a function of the excited state optimisation for the three systems shown in panels d-f). The curves on the excited state are colour-coded with the oscillator strengths. S_0 and S_1 refer to the ground and excited state energies, respectively. The right-side of the y-axis corresponds to C=O bond lengths (dashed black lines) for g) and h). In panel i), where there is a proton transfer on the excited state, the dashed blue lines correspond to the O-H distance along the SHB on the same distance scale.

Fig. 4b shows a scatter plot of the difference between the first excited state and ground-state energies and the corresponding oscillator strengths. The scatter plots were obtained over the course of the excited state optimisation. We observe that the L-pyro-amm system is characterised by the largest oscillator strengths compared to L-glu or L-pyro. One of the major structural changes that occurs upon excitation is an increase in the C=O bond length similar to observations in previous studies (38–40). Fig. 4c shows the oscillator strengths as a function of the C=O bond, that increases in length by about 0.1 Angstroms.

Although all three glutamine-related structures display similar trends on the excited state, what makes L-pyro-amm unique is the presence of the SHB. Fig. 4b and c show that L-pyro-amm displays the largest oscillator strength peaking at approximately 3.5eV consistent with our experimental findings. Although L-glu also has a peak at around 3.5 eV it is much weaker

than the one of L-pyro-amm. We thus decided to focus on a series of excited state optimisations for various clusters of L-pyro-amm. In particular, we focused on the proton transfer in the ground state as seen in the double-well potential and on the proximity of the ammonium ion to the short hydrogen bond region, all shown to play an important role in the optical properties of L-pyro-amm so far. We thus constructed three clusters with different initial conditions, which are shown in the top panels of Fig. 4d-f. The clusters shown in Fig. 4d and 4e differ in the initial positions of the protons along the SHB. As seen in Fig. 4a, the ground-state simulations of L-pyro-amm present a double-well potential in the finite temperature simulations. We thus moved the proton along one of the SHBs, constrained the geometry in the ground-state which yielded the cluster shown in Fig. 4e. In Fig. 4f, the ammonium ion is placed closer to the SHB region.

The circled regions in Fig. 4d-f highlight the main regions, where the nuclear degrees of freedom respond in the excited state. The bottom panels show a zoomed in image of the changes that occur upon relaxation, for which the solid and transparent structures correspond to geometries obtained before and after the relaxation on the excited state, respectively. The bottom panel of Fig. 4d illustrates that the change involves the C=O stretch and deplanarisation of the peptide dihedral angle of the ring. The corresponding evolution of the first excited state and ground state energies as a function of optimisation is shown in Fig. 4g. The relaxation on the excited state narrows the energy difference between the ground and excited state to approximately 3.5eV which is consistent with the experiments.

The top panels of Fig. 4e and f involve two other initial conditions for which the proton vibrations and the proximity of the ammonium ion to the SHB are altered. We see in Fig. 4h and i that the magnitude of the oscillator strength of L-pyro-amm in the excited state is sensitive to these new initial conditions. Transferring the protons on the ground-state (see dotted black rectangles in Fig. 4d-f), significantly changes the oscillator strengths as seen in the comparison between Fig. 4g-i, across the entire relaxation process in the first excited state while still maintaining the C=O stretch and deplanarisation as seen in the bottom panel of Fig. 3e. In the case of Fig. 4f, besides the reorganisation involving the C=O stretch, we also observe a proton transfer along the SHB. This is seen in Fig. 4i, which shows the O-H stretch (dashed-blue curve) along the SHB during the optimisation which increases from

1.26 to 1.32 Angstroms. The zoomed in plot in the bottom panel of Fig. 4f shows this subtle change in the proton transfer coordinate. Thus, the evolution of the excited state in L-pyro-amm into the region displaying visible fluorescence appears to involve a collective reorganisation of various vibrational modes forming the hydrogen bond network with the SHB playing a particular role, as it leads to the brightest structure, also confirmed experimentally. These findings are also consistent with some recent theoretical work showing the importance of proton transfer in self-assembled peptides made of aromatic phenyl-alanine amino acids (41). However, the latter structure features an aromatic amino acid, whereas we observe similar optical properties in non-aromatic structures.

Conclusions

The experimental and theoretical findings presented here, elucidate a rather complex molecular mechanism associated with the non-aromatic intrinsic fluorescence in protein-like structures. In the case of L-glutamine, a chemical reaction creates a newly formed structure involving a cyclised pyroglutamic acid ring. This new structure features absorption in the UV and emission in the visible range very similar to the chemically distinct amyloid fibrils (4, 5, 11, 12, 41–43).

The structural chromophore responsible for the optical properties in this new protein-related structure arises from a hydrogen bond network characterised by specific vibrational fluctuations on the excited state. The presence of strong hydrogen bonds along which proton transfer occurs and secondly, specific ionic interactions in close proximity, such as involving the ammonium ion, affect the optical properties. Although the fluorescence observed in these systems is much weaker than those compared to conventional fluorophores, the physical and chemical properties of the hydrogen bond network reported here maybe a generic feature across many other peptide structures.

In summary, we show here that there are several vibrational modes in biological structures that contribute to fluorescence properties as shown by excited state simulations and confirmed experimentally. We further show that the protein environment strongly contributes to the optical activity of the protein like structure and that the SHB significantly

enhances the fluorescence signal detected, potentially by inhibiting a conical intersection in the excited state as discussed previously (5). Interestingly, SHBs have recently been observed in different biological systems which have long been associated with either intrinsic fluorescence, such as NADP/NAD (44), FAD/FMN (45), the light-sensing chromophore in photoactive yellow protein (46), or in the active site of many enzymes, such as hydrolases and oxidoreductases (47, 48), many of which consist of a highly complex H-bond structure similar to amyloid proteins. Our findings may thus further the design of novel optically active biomaterial for applications in optical sensing or the design of novel biocompatible catalysts.

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Author Contributions

*A.D.S and M.N.Q contributed equally. A.D.S, prepared samples for all experimental data. P.J.W. performed SEM experiments. A.D.B and M.T.R performed XRD measurements and analysed data. M.T.R. performed THz experiments and DFT-THz calculations and analysed data. A.D.S. performed excitation and emission measurements and analysed data. A.D.S and S.T.J. performed absorption measurements and analysed data. M.T.R and E.M.K performed TD-DFT cluster calculations. M.N.Q, E.P, L.G, R.G and A.H performed AIMD, Periodic TD-DFT Excited State Calculations and Periodic Structure Geometry Optimisation calculations. A.D.S, M.N.Q, M.T.R, S.T.J, L.G, J.A.Z, D.C, A.H and G.S.K.S contributed to manuscript writing. All authors have given approval to the final version of the manuscript.

Notes

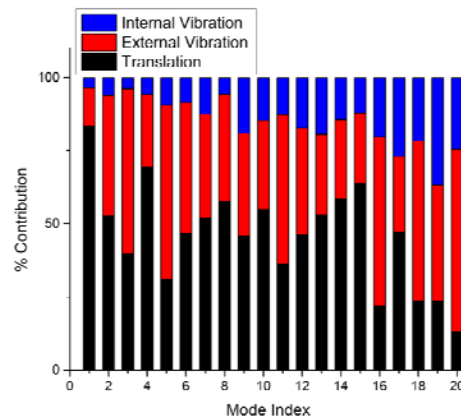
The authors declare no competing financial interest.

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Supplementary data

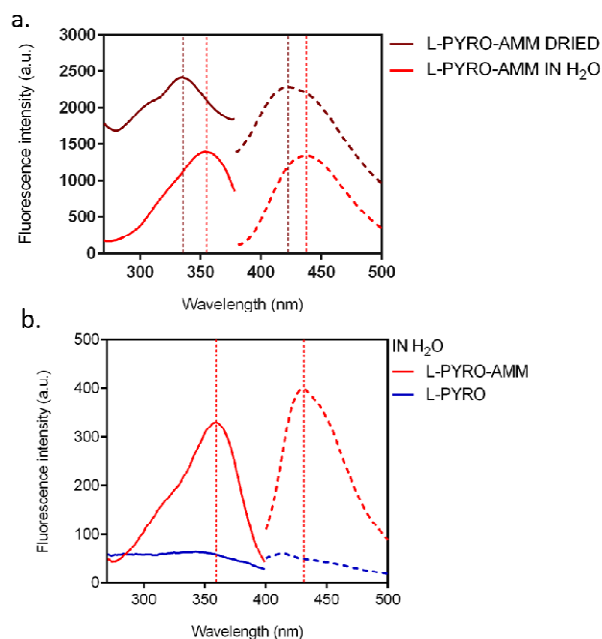


Mode Number	Frequency (THz)	Intensity (km mol ⁻¹)	Assignment
1	0.955	0.22	Antisymmetric translation in b
2	1.215	0.71	Coupled translation in a and rotation about bc
3	1.325	1.52	external rotation about b
4	1.450	0.36	Asymmetric translation in c
5	1.554	4.37	External rotation about a
6	1.576	1.76	Translation in a and external asymmetric rotation about H-bond coordinate
7	1.805	2.28	External asymmetric rotation of dimer pairs
8	1.929	2.11	External symmetric rotation of dimer pairs
9	1.950	1.28	Asymmetric rotation about H-bond coordinate, ammonium translational motion
10	2.113	3.53	Translation and rotation (breathing) around ammonium cation
11	2.208	2.5	Symmetric external rotation of entire formula units
12	2.404	2.37	In phase external rotation perpendicular to H-bond coordinate
13	2.502	0.45	Out of phase external rotation perpendicular to H-bond coordinate
14	2.564	0.73	External out of phase rotation of H-bonded chains about b
15	2.565	8.91	Out of phase external rotation of individual pyroglutamic molecules with translation of ammonium cation
16	2.654	18.09	External vibration coupled with torsion of the COOH group
17	2.679	0.29	External rotation and torsion of pyroglutamic ring
18	2.776	4.84	External rotation and torsion of ring and carboxyl group
19	2.988	2.68	External rotation and translation of ammonium

Supplementary Figure 1. Full spectral assignment of THz data.

The top chart shows the contribution to each IR-active mode including external translations and hindered rotations, and internal vibrational motions (i.e. torsions), while the bottom table lists the detailed assignment for each mode.

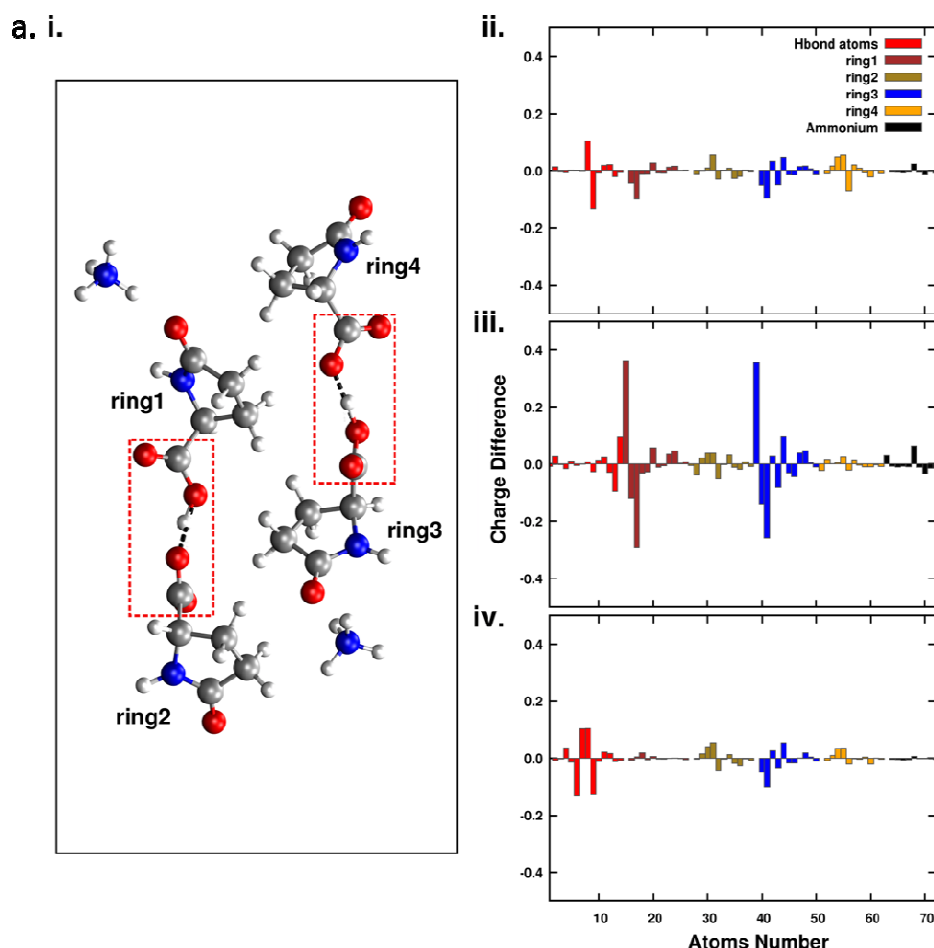
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543

544 **Supplementary Figure 2. L-pyro-amm has blue shifted fluorescence when dried and**
 545 **displays higher fluorescence intensity than L-pyro.**

546 (a) The excitation peak of L-pyro-amm when dried (solid dark red line) is blue shifted with a
 547 peak maximum ~ 340 nm compared to L-pyro-amm in H₂O (solid red line) which has a peak
 548 maximum ~ 360 nm. The emission peak of L-pyro-amm when dried (dashed dark red line) is
 549 also blue shifted, with a peak maximum ~ 420 nm, compared to L-pyro-amm in H₂O (dashed
 550 red line) with a peak maximum ~ 430 nm. (b) 1 M L-glu and 1 M L-pyro (blue) were
 551 incubated in H₂O for 8 days at 65°C. After 9 days, the L-glu had converted into the L-pyro-
 552 amm structure (red). L-pyro-amm has a clear excitation peak maximum at ~360 nm and
 553 emission peak maximum at ~ 430 nm, while L-pyro (blue), although not completely dark, has
 554 no clear excitation or emission peak.



Supplementary Figure 3. The optical properties of L-pyro-amm are sensitive to the environment and involves the electronic response of the entire structure.

(a)(i) L-pyro-amm cluster used to examine the sensitivity of the optical response on different parts of the cluster upon moving different protons. (ii) Charge differences between the ground and excited state are computed using restrained electrostatic potential atomic partial charges (RESP) for the cluster shown in i). Note, the electronic response involves all the atoms of the cluster. (iii) The two protons in the rectangle regions are displaced to be in the centre of the hydrogen bond and the charge differences are then computed. As illustrated, the proton displacement leads to a larger change in magnitude of the charges. (iv) The charge differences are computed for another nuclear configuration for which the protons are kept fixed but the O—O distance is increased from 2.45 to 3.2 Angstroms. The charge differences obtained here are quite similar to the original condition shown in (ii).

References:

1. A. Shukla, S. Mukherjee, S. Sharma, V. Agrawal, K. V. R. Kishan, P. Guptasarma, A novel UV laser-induced visible blue radiation from protein crystals and aggregates: Scattering artifacts or fluorescence transitions of peptide electrons delocalized through hydrogen bonding? *Arch. Biochem. Biophys.* **428**, 144–153 (2004).
2. G. Rosenman, N. Amdursky, M. Molotskii, D. Aronov, L. Adler-Abramovich, E. Gazit, Blue luminescence based on quantum confinement at peptide nanotubes. *Nano Lett.* **9**, 3111–3115 (2009).
3. F. T. Chan, D. Pinotsi, G. S. K. Schierle, C. F. Kaminski, in *Bio-nanoimaging: Protein Misfolding and Aggregation* (Elsevier, 2013; <http://linkinghub.elsevier.com/retrieve/pii/B9780123944313000134>), pp. 147–155.
4. F. T. S. Chan, G. S. Kaminski Schierle, J. R. Kumita, C. W. Bertoncini, C. M. Dobson, C. F. Kaminski, C. V Robinson, C. M. Dobson, J. Beard, P. Das, K. Jansen, M. DeLucia, W.-L. Lin, G. Dolios, R. Wang, C. B. Eckman, D. W. Dickson, M. Hutton, J. Hardy, T. Golde, Protein amyloids develop an intrinsic fluorescence signature during aggregation. *Analyst.* **138**, 2156 (2013).
5. D. Pinotsi, L. Grisanti, P. Mahou, R. Gebauer, C. F. Kaminski, A. Hassanali, G. S. Kaminski Schierle, Proton Transfer and Structure-Specific Fluorescence in Hydrogen Bond-Rich Protein Structures. *J. Am. Chem. Soc.* **138**, 3046–3057 (2016).
6. J. Pansieri, V. Josserand, S. J. Lee, A. Rongier, D. Imbert, M. M. Sallanon, E. Kövari, T. G. Dane, C. Vendrely, O. Chaix-Pluchery, M. Guidetti, J. Vollaie, A. Fertin, Y. Usson, P. Rannou, J. L. Coll, C. Marquette, V. Forge, Ultraviolet–visible–near-infrared optical properties of amyloid fibrils shed light on amyloidogenesis. *Nat. Photonics.* **13**, 473–479 (2019).
7. L. Adler-Abramovich, E. Gazit, Correction: The physical properties of supramolecular peptide assemblies: From building block association to technological applications. *Chem. Soc. Rev.* **43**, 7236–7236 (2014).
8. L. Adler-Abramovich, M. Reches, V. L. Sedman, S. Allen, S. J. B. Tendler, E. Gazit, Thermal and chemical stability of diphenylalanine peptide nanotubes: Implications for nanotechnological applications. *Langmuir.* **22**, 1313–1320 (2006).
9. K. Tao, P. Makam, R. Aizen, E. Gazit, Self-assembling peptide semiconductors. *Science (80-.).* **358** (2017), p. eaam9756.

- 602 10. J. M. Andresen, J. Gayán, L. Djoussé, S. Roberts, D. Brocklebank, S. S. Cherny, L. R.
603 Cardon, J. F. Gusella, M. E. Macdonald, R. H. Myers, D. E. Housman, N. S. Wexler, J.
604 Lorimer, J. Porter, F. Gomez, C. Moskowitz, K. P. Gerstenhaber, E. Shackell, K. Marder,
605 G. Penchaszadeh, S. A. Roberts, A. Brickman, D. Brocklebank, J. Gray, S. R. Dlouhy, S.
606 Wiktorski, M. E. Hodes, P. M. Conneally, J. B. Penney, J. F. Gusella, J. H. Cha, M.
607 Irizarry, D. Rosas, S. Hersch, Z. Hollingsworth, A. B. Young, D. E. Housman, M. M. de
608 Young, E. Bonilla, T. Stillings, A. Negrette, S. R. Snodgrass, M. D. Martinez-Jaurrieta,
609 M. A. Ramos-Arroyoh, J. Bickham, J. S. Ramos, F. Marshall, I. Shoulson, G. J. Rey, A.
610 Feigin, N. Arnheim, A. Acevedo-Cruz, L. Acosta, J. Alvir, K. Fischbeck, L. M. Thompson,
611 A. Young, L. Dure, C. J. O'Brien, J. Paulsen, S. P. Moran, D. Krch, P. Hogarth, D. S.
612 Higgins, B. Landwehrmeyer, M. R. Hayden, E. W. Almqvist, R. R. Brinkman, O.
613 Suchowersky, A. Durr, C. C. Dodé, F. Squitieri, P. J. Morrison, M. Nance, C. A. Ross, R.
614 L. Margolis, A. Rosenblatt, G. T. Estrella, D. M. Cabrero, R. J. A. Trent, E. McCusker, A.
615 Novelletto, M. Frontali, J. S. Paulsen, R. Jones, A. Zanko, T. Ashizawa, A. Lazzarini, J. L.
616 Li, V. C. Wheeler, A. L. Russ, G. Xu, J. S. Mysore, T. Gillis, M. Hakky, L. A. Cupples, M.
617 Saint-Hilaire, S. M. Hersch, The relationship between CAG repeat length and age of
618 onset differs for Huntington's disease patients with juvenile onset or adult onset.
619 *Ann. Hum. Genet.* **71**, 295–301 (2007).
- 620 11. D. Pinotsi, A. K. Buell, C. M. Dobson, G. S. Kaminski Schierle, C. F. Kaminski, A Label-
621 Free, Quantitative Assay of Amyloid Fibril Growth Based on Intrinsic Fluorescence.
622 *ChemBioChem.* **14**, 846–850 (2013).
- 623 12. T. N. Tikhonova, N. R. Rovnyagina, A. Y. Zhrebker, N. N. Sluchanko, A. A. Rubekina, A.
624 S. Orekhov, E. N. Nikolaev, V. V. Fadeev, V. N. Uversky, E. A. Shirshin, Dissection of the
625 deep-blue autofluorescence changes accompanying amyloid fibrillation. *Arch.*
626 *Biochem. Biophys.* **651**, 13–20 (2018).
- 627 13. H. Mori, K. Takio, M. Ogawara, D. J. Selkoe, Mass spectrometry of purified amyloid β
628 protein in Alzheimer's disease. *J. Biol. Chem.* **267**, 17082–17086 (1992).
- 629 14. M. Fändrich, C. M. Dobson, The behaviour of polyamino acids reveals an inverse side
630 chain effect in amyloid structure formation. *EMBO J.* **21**, 5682–90 (2002).
- 631 15. R. Dovesi, A. Erba, R. Orlando, C. M. Zicovich-Wilson, B. Civalleri, L. Maschio, M. Rérat,
632 S. Casassa, J. Baima, S. Salustro, B. Kirtman, Quantum-mechanical condensed matter
633 simulations with CRYSTAL. *Wiley Interdiscip. Rev. Comput. Mol. Sci.* **8**, e1360 (2018).

- 634 16. P. Giannozzi, O. Andreussi, T. Brumme, O. Bunau, M. Buongiorno Nardelli, M.
635 Calandra, R. Car, C. Cavazzoni, D. Ceresoli, M. Cococcioni, N. Colonna, I. Carnimeo, A.
636 Dal Corso, S. De Gironcoli, P. Delugas, R. A. Distasio, A. Ferretti, A. Floris, G. Fratesi, G.
637 Fugallo, R. Gebauer, U. Gerstmann, F. Giustino, T. Gorni, J. Jia, M. Kawamura, H. Y. Ko,
638 A. Kokalj, E. Küçükbenli, M. Lazzeri, M. Marsili, N. Marzari, F. Mauri, N. L. Nguyen, H.
639 V. Nguyen, A. Otero-De-La-Roza, L. Paulatto, S. Poncé, D. Rocca, R. Sabatini, B. Santra,
640 M. Schlipf, A. P. Seitsonen, A. Smogunov, I. Timrov, T. Thonhauser, P. Umari, N. Vast,
641 X. Wu, S. Baroni, Advanced capabilities for materials modelling with Quantum
642 ESPRESSO. *J. Phys. Condens. Matter.* **29** (2017), doi:10.1088/1361-648X/aa8f79.
- 643 17. M. T. Ruggiero, J. Gooch, J. Zubieta, T. M. Korter, Evaluation of Range-Corrected
644 Density Functionals for the Simulation of Pyridinium-Containing Molecular Crystals. *J.*
645 *Phys. Chem. A.* **120**, 939–947 (2016).
- 646 18. J. Da Chai, M. Head-Gordon, Long-range corrected hybrid density functionals with
647 damped atom-atom dispersion corrections. *Phys. Chem. Chem. Phys.* **10**, 6615–6620
648 (2008).
- 649 19. D. Rocca, R. Gebauer, Y. Saad, S. Baroni, Turbo charging time-dependent density-
650 functional theory with Lanczos chains. *J. Chem. Phys.* **128** (2008),
651 doi:10.1063/1.2899649.
- 652 20. N. Troullier, J. L. Martins, Efficient pseudopotentials for plane-wave calculations.
653 *Phys. Rev. B.* **43**, 1993–2006 (1991).
- 654 21. A. D. Becke, Density-functional thermochemistry. III. The role of exact exchange. *J.*
655 *Chem. Phys.* **98**, 5648–5652 (1993).
- 656 22. J. D. Head, M. C. Zerner, A Broyden-Fletcher-Goldfarb-Shanno optimization procedure
657 for molecular geometries. *Chem. Phys. Lett.* **122**, 264–270 (1985).
- 658 23. J. VandeVondele, M. Krack, F. Mohamed, M. Parrinello, T. Chassaing, J. Hutter,
659 Quickstep: Fast and accurate density functional calculations using a mixed Gaussian
660 and plane waves approach. *Comput. Phys. Commun.* **167**, 103–128 (2005).
- 661 24. S. Goedecker, M. Teter, Separable dual-space Gaussian pseudopotentials. *Phys. Rev.*
662 *B - Condens. Matter Mater. Phys.* **54**, 1703–1710 (1996).
- 663 25. A. D. Becke, Density-functional exchange-energy approximation with correct
664 asymptotic behavior. *Phys. Rev. A.* **38**, 3098–3100 (1988).
- 665 26. S. Grimme, J. Antony, S. Ehrlich, H. Krieg, A consistent and accurate *ab initio*

- 666 parametrization of density functional dispersion correction (DFT-D) for the 94
- 667 elements H-Pu. *J. Chem. Phys.* **132**, 154104 (2010).
- 668 27. G. Bussi, D. Donadio, M. Parrinello, Canonical sampling through velocity rescaling. *J.*
- 669 *Chem. Phys.* **126** (2007), doi:10.1063/1.2408420.
- 670 28. T. Yanai, D. P. Tew, N. C. Handy, A new hybrid exchange-correlation functional using
- 671 the Coulomb-attenuating method (CAM-B3LYP). *Chem. Phys. Lett.* **393**, 51–57 (2004).
- 672 29. K. Jong, Y. Taghipour Azar, L. Grisanti, A. D. Stephens, S. T. E. Jones, D. Credgington, G.
- 673 S. Kaminski Schierle, A. A. Hassanali, Low Energy Optical Excitations as an Indicator of
- 674 Structural Changes Initiated at the Termini of Amyloid Protein. *Phys. Chem. Chem.*
- 675 *Phys.* (2019), doi:10.1039/c9cp04648h.
- 676 30. L. Grisanti, D. Pinotsi, R. Gebauer, G. S. Kaminski Schierle, A. A. Hassanali, A
- 677 computational study on how structure influences the optical properties in model
- 678 crystal structures of amyloid fibrils. *Phys. Chem. Chem. Phys.* **19**, 4030–4040 (2017).
- 679 31. E. P. J. Parrott, J. A. Zeitler, Terahertz time-domain and low-frequency Raman
- 680 spectroscopy of organic materials. *Appl. Spectrosc.* **69**, 1–25 (2015).
- 681 32. Y. K. Law, A. A. Hassanali, Role of Quantum Vibrations on the Structural, Electronic,
- 682 and Optical Properties of 9-Methylguanine. *J. Phys. Chem. A.* **119**, 10816–10827
- 683 (2015).
- 684 33. S. Sappati, A. Hassanali, R. Gebauer, P. Ghosh, Nuclear quantum effects in a
- 685 HIV/cancer inhibitor: The case of ellipticine. *J. Chem. Phys.* **145** (2016),
- 686 doi:10.1063/1.4968046.
- 687 34. Y. K. Law, A. A. Hassanali, The importance of nuclear quantum effects in spectral line
- 688 broadening of optical spectra and electrostatic properties in aromatic chromophores.
- 689 *J. Chem. Phys.* **148** (2018), doi:10.1063/1.5005056.
- 690 35. D. Marx, Erratum: Proton transfer 200 years after von grotthuss: Insights from Ab
- 691 initio simulations (ChemPhysChem (2006) 7 (1848-1810)). *ChemPhysChem.* **8**, 209–
- 692 210 (2007).
- 693 36. S. Prasad, I. Mandal, S. Singh, A. Paul, B. Mandal, R. Venkatramani, R. Swaminathan,
- 694 Near UV-Visible electronic absorption originating from charged amino acids in a
- 695 monomeric protein. *Chem. Sci.* **8**, 5416–5433 (2017).
- 696 37. I. Mandal, S. Paul, R. Venkatramani, Optical backbone-sidechain charge transfer
- 697 transitions in proteins sensitive to secondary structure and modifications. *Faraday*

- 698 *Discuss.* **207**, 115–135 (2018).
- 699 38. M. Mališ, Y. Loquais, E. Gloaguen, H. S. Biswal, F. Piuze, B. Tardivel, V. Brenner, M.
700 Broquier, C. Juvet, M. Mons, N. D. S. Došlić, I. Ljubić, Unraveling the mechanisms of
701 nonradiative deactivation in model peptides following photoexcitation of a
702 phenylalanine residue. *J. Am. Chem. Soc.* **134**, 20340–20351 (2012).
- 703 39. B. P. Fingerhut, K. E. Dorfman, S. Mukamel, Monitoring nonadiabatic dynamics of the
704 RNA base uracil by UV pump-IR probe spectroscopy. *J. Phys. Chem. Lett.* **4**, 1933–
705 1942 (2013).
- 706 40. M. Sapunar, T. Ayari, N. Došlić, Comparative study of the photodynamics of
707 malonaldehyde and acetylacetone. *Chem. Phys.* **515**, 622–627 (2018).
- 708 41. S. K. Joseph, N. Kuritz, E. Yahel, N. Lapshina, G. Rosenman, A. Natan, Proton-Transfer-
709 Induced Fluorescence in Self-Assembled Short Peptides. *J. Phys. Chem. A.* **123**, 1758–
710 1765 (2019).
- 711 42. A. Handelman, N. Kuritz, A. Natan, G. Rosenman, Reconstructive Phase Transition in
712 Ultrashort Peptide Nanostructures and Induced Visible Photoluminescence.
713 *Langmuir*. **32** (2016), pp. 2847–2862.
- 714 43. L. L. Del Mercato, P. P. Pompa, G. Maruccio, A. Della Torre, S. Sabella, A. M.
715 Tamburro, R. Cingolani, R. Rinaldi, Charge transport and intrinsic fluorescence in
716 amyloid-like fibrils. *Proc Natl Acad Sci U S A.* **104**, 18019–18024 (2007).
- 717 44. A. Stavrinos, E. C. Tatsis, L. Caputi, E. Foureau, C. E. M. Stevenson, D. M. Lawson, V.
718 Courdavault, S. E. O'Connor, Structural investigation of heteroyohimbine alkaloid
719 synthesis reveals active site elements that control stereoselectivity. *Nat. Commun.* **7**
720 (2016), doi:10.1038/ncomms12116.
- 721 45. F. Forneris, D. P. H. M. Heuts, M. Delvecchio, S. Roviola, M. W. Fraaije, A. Mattevi,
722 Structural analysis of the catalytic mechanism and stereoselectivity in *Streptomyces*
723 *coelicolor* alditol oxidase. *Biochemistry.* **47**, 978–985 (2008).
- 724 46. S. Yamaguchi, H. Kamikubo, K. Kurihara, R. Kuroki, N. Niimura, N. Shimizu, Y.
725 Yamazaki, M. Kataoka, Low-barrier hydrogen bond in photoactive yellow protein.
726 *Proc. Natl. Acad. Sci. U. S. A.* **106**, 440–444 (2009).
- 727 47. W. W. Cleland, P. A. Frey, J. A. Gerlt, The low barrier hydrogen bond in enzymatic
728 catalysis. *J. Biol. Chem.* **273** (1998), pp. 25529–25532.
- 729 48. S. Zhou, L. Wang, Unraveling the structural and chemical features of biological short

730 hydrogen bonds. *Chem. Sci.* **10**, 7734–7745 (2019).

731