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Theme: *Dietary muscle proteins for stage of life*

Dietary meat and protection against sarcopenia

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Abstract

Sarcopenia describes the age-related loss of skeletal muscle mass and associated muscle weakness. Sarcopenia is a major global health problem given that the number and proportion of older people in the population is escalating worldwide and represent the fastest growing segment of society. The loss of muscle mass compromises physical capacity, increases susceptibility to falls, and impacts on an individual's functional independence and quality of life. Tackling sarcopenia sensibly and effectively will identify strategies that will enable older adults to age well and age productively. The underlying causes of sarcopenia are complex and multifactorial and will likely require combinatorial therapies to address its symptoms. Nutrition, particularly protein intake, is a more easily modifiable factor, especially when combined with structured (resistance) exercise programs. The relative success of protein feeding strategies for sarcopenia, is limited by a so-called anabolic resistance in older people. Meat contains essential amino acids and nutritive compounds of high quality, and even a moderate intake can increase muscle protein synthesis in older men and women. However, health risks have been identified with the consumption of different meats, with high intake of processed meats increasing the risk for cardiovascular disease and different cancers. Risks for fresh white and red meat are considerably less and modest consumption is encouraged as part of a healthy eating plan for many older adults to ensure adequate protein intake. Other nutritive strategies of relevance for sarcopenia involve fortifying the nutrient value of different meats. Studies on muscle cells and animal models of muscle wasting, have identified the therapeutic potential of the amino acid, glycine, to reduce inflammation, attenuate muscle atrophy, and re-sensitize muscle to anabolic stimuli. Glycine supplementation or feeding animal products with a high glycine content (e.g. gelatin), could represent simple and effective nutritional strategies as part of a suite of therapies to attenuate sarcopenia.

Sarcopenia – a global public health problem

Skeletal muscle is essential for life – not just as an organ for locomotion or breathing, but as an endocrine organ that contracts and releases different factors (‘myokines’) that signal between other organs and tissues (Whitham et al. 2018). Many muscle wasting diseases and conditions are associated with increased morbidity and mortality and therefore maintaining muscle mass can be critical for survival. Muscle mass is maintained through a balance between protein synthesis and protein degradation. Different factors can influence this balance ultimately to determine whether muscle mass is gained (muscle hypertrophy) or lost (muscle atrophy). These different intrinsic and extrinsic factors include nutrition, genetics, innervation, hormones, metabolism, inflammation, oxidative stress, exercise (physical activity) and diseases. These factors are implicated to some extent with the loss of muscle homeostasis with aging. ‘Sarcopenia’ describes this age-related muscle wasting mass and weakness. Although precise definitions continue to be controversial, the criteria for the clinical diagnosis of sarcopenia include the presence of low muscle mass accompanied by low muscle strength and/or low physical performance (Evans 2010; Reijnierse et al. 2016). Aging compromises skeletal muscle structure, metabolism and function, although generally there is a greater loss of muscle strength than mass and weakness appears more closely associated with the risk of disability and mortality (Evans 2015). It has been suggested that a more comprehensive definition of sarcopenia should also include measures of insulin resistance and basal metabolic rate and these measurements together with accurate assessments of muscle mass and strength would provide an index of the actual contribution of skeletal muscle to the age-associated risk of morbidity and mortality (Evans 2015).

Sarcopenia is an important global public health problem affecting especially the developed world where the number and proportion of older adults in the populations is escalating (Ortman et al. 2014). The significance of the problem is clear when one considers

that the proportion of adults aged over 60 years is expected to increase to 27% by 2050 and that 5-13% of older adults have low muscle mass (as high as 50% in those aged 80 years and older). In normal aging there is a 1% loss of muscle mass after 30 years of age, which tends to accelerate after 70 years of age (Morley et al. 2014; Baugreet et al. 2017). Frail elders who have lost significant muscle mass and strength often require assistance for accomplishing even the most basic tasks of independent living, and they are also at increased risk of serious injury from sudden falls and fractures. This imposes a significant but modifiable economic burden on healthcare services in most industrialized nations (Lynch 2011). There is clearly an urgent and unmet clinical need to better understand the mechanisms responsible for sarcopenia and to develop interventions that can attenuate, prevent, and possibly reverse sarcopenia.

Many of the contributing cellular and molecular mechanisms underlying age-related muscle wasting and weakness have been described in detail elsewhere (Lynch 2011; Clark & Fielding 2012; Argiles et al. 2015; Cohen et al. 2015; Hepple & Rice 2016; Sousa-Victor & Muñoz-Cánoves 2016). Sarcopenia is characterized by a slow, progressive decline in muscle quantity and quality attributed to the progressive atrophy and loss of individual muscle fibers associated with the loss of some motor units, and a reduction in muscle ‘quality’ because of alterations in muscle metabolism. Age-related changes in skeletal muscle structure and function are associated with a complex interaction of factors affecting neuromuscular transmission, motor unit remodeling and shifts in muscle fiber composition. There are age-related alterations in excitation-contraction coupling, increased generation of reactive oxygen species and myonuclear apoptosis, changes in muscle architecture including loss of muscle fibers and infiltration of fibrotic and other non-contractile material. There are also decreases in circulating levels of anabolic factors and hormones such as testosterone, growth hormone, and insulin-like growth factor-I, and higher systemic and local (muscle) levels of

inflammatory markers including tumor necrosis factor-alpha (TNF- α), interleukin-6 (IL-6) and C reactive protein (CRP) (Ryall et al. 2008; Schaap et al. 2009; Tintignac et al. 2015; Ogawa et al. 2016; Piasecki et al. 2016; Welch et al. 2018).

Anabolic resistance – implications for sarcopenia

The decrease in skeletal muscle mass with aging can be attributed to a chronic disturbance in the regulation of skeletal muscle protein turnover (Figure 1), leading to an imbalance between protein synthesis and protein breakdown (Koopman & van Loon 2009). Although it was originally assumed that healthy older people had decreased rates of basal muscle protein synthesis (Short et al. 2004), consideration of factors such as health status, habitual physical activity and dietary habits, led to the consensus that there is little or no difference in basal muscle protein synthesis rates between healthy young and old individuals (Cuthbertson et al. 2005; Katsanos et al. 2005; Symons et al. 2007). Since the rate of muscle loss with aging is less than two per cent per year, there is support for the hypothesis that basal fasting protein synthesis and/or breakdown rates are not substantially impaired with aging (Volpi et al. 2001, Rennie 2009, Phillips et al. 2017). In contrast, the protein synthetic response to anabolic stimuli such as protein ingestion and physical exercise is impaired in older individuals (Wall et al. 2015). As humans are under some degree of feeding- and exercise-induced stimulation of protein synthesis during most of the day, this so-called ‘anabolic resistance’ is primarily responsible for skeletal muscle atrophy during aging (Moore 2014; Murton 2015; Morton et al. 2018). The exact mechanisms responsible for anabolic resistance have yet to be established, but inactivity and chronic low-grade inflammation are strongly associated with physical decline in older humans (Fougere et al. 2017) and considered probable instigators of anabolic resistance (Ham et al. 2014a). Older adults are more likely to experience periods of inactivity through lifestyle changes or acute or chronic illnesses and progress from a state of healthy aging to frailty and loss of functional independence. Thus, the importance of

structured exercise (especially resistance training) and incidental activity during aging cannot be underestimated as an essential self-directed behavior that can help retain the capacity for a robust anabolic response to dietary protein (Symons et al. 2011; Morris & Jacques 2013; Moore 2014). In addition to age-related changes in physical activity contributing to nutritional deficiencies in older adults, other factors also need to be considered, including changes in cognitive function, alterations in taste and chemosensory acuity, impaired swallowing with difficulties in chewing because of associated problems with teeth and gums, and potential issues arising from the need to take medications for various health conditions (Baugreet et al. 2017).

We have demonstrated that during acute inflammation, the anabolic response to leucine is impaired (Ham et al. 2016). The failure of leucine to stimulate skeletal muscle protein synthesis, even at higher doses, has been also shown in rodent models of sepsis and endotoxemia (Lang & Frost 2004; 2005). These observations highlight the impact of inflammation on amino acid sensing and stimulation of protein synthesis and challenges the hypothesis that anabolic resistance can be overcome simply by providing more substrate (i.e. dietary protein or leucine) (Ham et al. 2014a). In addition to providing adequate amounts of protein, we propose that other nutrients or treatments that sensitize skeletal muscle to leucine have therapeutic potential for muscle wasting conditions.

Nutritional considerations for sarcopenia – importance of dietary protein

Protein intake is among the factors that regulate protein turnover. The amount and quality of protein is well-known to determine the protein synthetic response to meal ingestion. A dose-response study in healthy older men using whey proteins found that ingestion of 30-40 grams of protein stimulated protein synthesis more than smaller amounts (Pennings et al. 2011). Overall, these acute studies suggest that ensuring each meal contains this amount of protein would provide health benefits for this population. However, most older individuals consume

a breakfast containing relatively low protein content (Tieland et al. 2015) or skip breakfast altogether (Deshmukh-Taskar et al. 2013). Including a high-quality protein source at breakfast may be the easiest way to increase daily protein intake. Data from large cohort studies suggest that the loss of muscle mass in older men and women is attenuated in those that consume considerably more protein than the current Recommended Dietary Allowance (RDA, 0.8 g/kg/d) (Baum et al. 2016; Courtney-Martin et al. 2016). In fact, recommendations from the ESPEN Expert Group for healthy older people is that the diet should provide at least 1.0-1.2 g protein/kg body weight/day (Deutz et al. 2014).

In addition to the amount of protein, digestibility and quality also determine the anabolic response. Generally, ingestion of faster digestible proteins results in a more rapid increase in plasma amino acid concentration and a greater muscle protein synthetic response. Indeed, ingestion of a casein protein hydrolysate (i.e. predigested casein), accelerates protein digestion and absorption, enhances postprandial amino acid availability compared to its intact protein, and increases the skeletal muscle protein synthesis (Koopman et al. 2009). Whey protein stimulates protein synthesis to a greater extent than casein (Pennings et al. 2011). Digestibility of protein is not the only factor that differentiates whey and casein. The anabolic response to casein hydrolysate is markedly less than that of whey, whereas the digestion and absorption kinetics are similar (Pennings et al. 2011). The difference in protein synthesis following intake of whey and casein hydrolysate is mainly attributed to the difference in leucine content. Clearly, amino acid composition of dietary proteins plays an important regulatory role in postprandial muscle protein synthesis.

The quality of protein is mainly determined by its amino acid profile as different amino acids have different anabolic properties (Landi et al. 2016). For example, L-arginine and its metabolite L-citrulline are known to protect muscle cells from wasting by modulating protein synthesis (Ham et al. 2014c). Generally, essential amino acid availability is a key driver of

skeletal muscle protein synthesis and the branched-chain amino acid leucine plays a unique role in the modulation of skeletal muscle metabolism (Ham et al. 2014). Unlike any other amino acid, leucine directly modulates the mechanistic Target of Rapamycin Complex 1 (mTORC1), one on the main anabolic signaling pathways in skeletal muscle. Indeed, leucine administration has been demonstrated to increase protein synthesis in cell culture systems and mammals. In addition, studies in humans have clearly demonstrated that the acute anabolic response to a suboptimal dose of protein can be enhanced when the leucine content of the protein bolus is increased (Wall et al. 2013). As a result, studies have been performed to assess whether longer-term leucine supplementation can enhance muscle mass. Initial studies using modest doses of leucine (2.5 g leucine/meal) did not show any improvements in muscle mass (Verhoeven et al. 2009; Leenders et al. 2011). More recent work from the Phillips laboratory suggests that higher doses of leucine (5g leucine/meal) can chronically increase protein synthesis in older males and females potentially reducing muscle wasting in that population (Murphy et al. 2016; Devries et al. 2018).

Meat as an excellent source of protein

Meat contains a large quantity of essential amino acids and nutritive factors of high quality and availability, including minerals like iron and zinc, and a variety of vitamins, especially B group vitamins (Rondanelli et al. 2015). Even a moderate intake of lean meat has been shown to increase protein synthesis in both young and old of both sexes (Symons et al. 2007; WHO 2007) and synergistic effects between meat intake and resistance exercise to increase muscle mass in older people have been reported (Daly et al. 2014). Meat also contains other compounds that can influence protein metabolism, including creatine, carnitine, carnosine, and conjugated linoleic acid (Rahman et al. 2009; Tarnopolsky & Safdar 2008; Moon et al. 2013; Young et al. 2013; Artioli et al. 2018). Some authors have even touted a diet for preventing sarcopenia that proposes eating meat 4-5 times a week; including white meat

twice per week, lean red meat less than twice per week, and processed meat less than once per week (Rondanelli et al. 2015).

Some studies have shown high consumption of meat products (especially processed meats) are linked with unfavorable health outcomes and increased risk for cardiovascular diseases, cognitive impairments, and some cancers, particularly colorectal cancer (Kouvari et al. 2016; Tabung et al. 2017). Other studies have disputed these findings (Binnie et al. 2014; McNeill 2014). Processed meat products are typically red meats that have been cured, salted or smoked (e.g. ham or bacon) to improve their durability, color and taste, and often contain high amounts of minced fatty tissue (e.g. sausages) (Rohrmann & Linseisen 2016). High consumption of processed meats may lead to an increased intake of saturated fats, cholesterol, salt, nitrite, and heme iron. Multiple (large cohort) studies have linked processed meat consumption and risk of chronic diseases such as cardiovascular disease, diabetes, and different cancers (Cascella et al. 2018; Quintana Pacheco et al. 2018).

Abete and colleagues (2014) conducted a meta-analysis of thirteen cohort studies to examine the association between consumption of meat (total, red, white and processed) and all-cause, cardiovascular disease and ischemic heart disease mortality. They found those in the highest category of processed meat consumption had 22% and 18% higher risk of mortality from any cause and cardiovascular disease, respectively. Red meat consumption was associated with a 16% higher risk of cardiovascular disease mortality, but there was no similar association for total and white meat consumption. While acknowledging the large heterogeneity in their data analyses they concluded that processed meat consumption could increase the risk of mortality from any cause and cardiovascular disease, and red meat consumption was positively but weakly associated with cardiovascular disease mortality (Abete et al. 2014). More moderate meat consumption within healthy adult populations may have less influence on morbidity or

mortality and some studies have suggested meat intake be reduced to a maximum of 70 grams per day (Millward & Garnett 2010).

In the context of sarcopenia, where reduced overall protein intake has considerable health implications for older people, attention is focused on increasing protein intake through nutrient-rich meats, fish and vegetable sources, and through supplements and fortified foods. The nutrient density of meat proteins has considerable potential to ameliorate sarcopenia (Phillips 2012). Considerations for meat consumption include the method of preparation and cooking since this influences the digestibility of the meat, amino acid absorption kinetics and subsequent stimulation of muscle protein synthesis. This is important for older individuals who generally experience a reduced food-chewing efficiency. Studies have assessed the effect of meat texture on dietary protein digestion rate, amino acid availability, and subsequent postprandial protein balance in older men. Pennings and colleagues (2013) demonstrated that ingestion of minced meat compared with steak resulted in a more rapid absorption of amino acids, leading to increased availability of amino acids in the circulation. Although no differences were observed in postprandial muscle protein synthesis, whole-body net protein accretion was significantly greater after minced meat intake. Similarly, consuming rare meat leads to less pronounced rises in plasma amino acid concentration in older individuals than consuming meat that has been cooked well-done (Buffière et al. 2017). Clearly, meat processing and cooking methods also affect postprandial amino acid availability and subsequent protein turnover.

Other meat products to support protein turnover

Meat is clearly a good source of amino acids and effective in stimulating skeletal muscle protein synthesis, and important for the maintenance of skeletal muscle mass. The anabolic resistance in older individuals may, however, necessitate other interventions that can sensitize the muscle for amino acids to support skeletal muscle protein turnover. It has been suggested

that other animal products may be valuable modulators of protein metabolism. Gelatin, a food derived from animal collagen, has been proposed to affect collagen synthesis with implications for injury prevention and tissue repair (Shaw et al. 2017). Gelatin has a very different amino acid profile than meat (Symons et al. 2007) or milk protein (Pennings et al. 2011) and is rich in glycine (23%), proline (13%) and hydroxyproline (11%) (Igase et al. 2018). Indeed, intake of 15 grams of gelatin rapidly increases plasma availability of glycine (2.2-fold) and proline (by 55%) (Shaw et al. 2017). As studies have demonstrated that muscle glycine levels are reduced in mouse models of diabetes (Giesbertz et al. 2015) aging (Houtkooper et al. 2011), and in muscle samples from frail older individuals (Fazelzadeh et al. 2016), we suggest that providing additional glycine could improve muscle metabolism during conditions of stress.

Glycine modulates cellular homeostasis via a receptor-mediated response and via its intracellular metabolism (Koopman et al. 2017). Glycine can directly activate glycine-gated chloride (Cl^-) channels (GlyR) expressed in inflammatory cells such as macrophages, thereby normalizing $[\text{Ca}^{2+}]$ and reducing pro-inflammatory cytokine production. In addition, glycine can deliver carbon units to the folate cycle, thereby modulating the production of cellular components (such as nucleotides and phospholipids) and affecting the balance between oxidized (GSSG) and reduced glutathione (GSH), a crucial modulator of cellular redox status. In a series of studies in mice to assess whether glycine administration protected muscles from wasting during inflammatory conditions, we showed glycine administration (1g/kg/day for 3 weeks) attenuated the loss of muscle mass and strength by 50% in a mouse model of cancer cachexia and attenuated the cancer-induced stimulation of muscle inflammation, macrophage infiltration and the production of reactive oxygen species (Ham et al. 2014b). Glycine also prevented superoxide production and maintained the protein synthetic response to leucine in an *in vivo* model of acute inflammation (lipopolysaccharide injections) in mice (Ham et al.

2016). Furthermore, glycine feeding (1g/kg/day) during calorie restriction, which is characterized by systemic inflammation, attenuated muscle atrophy in mice (Caldow et al. 2016; Ham et al. 2016). Combined, our data clearly highlight how glycine can effectively reduce inflammation, attenuate muscle loss during various condition, and re-sensitize skeletal muscle to anabolic stimuli. Therefore, animal products high in glycine, such as gelatin, could be simple and effective dietary supplements with anti-inflammatory actions that attenuate muscle wasting (Figure 1).

Conclusion

Advances in science, medicine and biomedical engineering (among other disciplines) have enabled humans to live longer lives, especially in developed countries. Living longer, however, does not necessarily mean living better, with many more older individuals presenting with diseases and conditions that compromise quality of life. Some of the most serious consequences of aging are its effects on skeletal muscle, with progressive age-related muscle wasting and weakness leading to a reduction in the ability to perform tasks of daily living, decreasing functional independence and having a devastating impact on quality of life. Addressing the underlying causes of sarcopenia and treating its consequences, represents a significant unmet clinical need. Possible interventions include pharmacologic and nutritional strategies and structured physical activity, especially resistance training. The relative success of nutritional (especially protein feeding) strategies for sarcopenia, is limited by the well-described phenomenon of ‘anabolic resistance’.

Meat contains essential amino acids and nutritive vitamins, minerals and other compounds of high quality, and a moderate intake of lean meat can increase protein synthesis in older men and women with synergistic effects when combined with resistance exercise training.

However, health risks have been identified with the consumption of different meats, with high intake of processed meats increasing the risk for cardiovascular disease and different cancers. The risks for fresh white and red meat are considerably less and modest consumption is encouraged for many older adults to ensure adequate protein intake. Fortifying meats to enhance their nutrient value also has relevance for sarcopenia. In studies on muscle cells and animal models of muscle wasting, the amino acid, glycine, reduced inflammation, attenuated muscle atrophy, and re-sensitized skeletal muscle to anabolic stimuli. Glycine feeding or animal products with a high glycine content (such as gelatin), could represent simple and effective nutritional strategies for attenuating age-related muscle wasting and weakness.

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References

- Abete, I., Romaguera, D., Vieira, A.R., Lopez de Munain, A., & Norat, T. (2014). Association between total, processed, red and white meat consumption and all-cause, CVD and IHD mortality: a meta-analysis of cohort studies. *British Journal of Nutrition* 112(5), 762-775.
- Argilés, J.M., Busquets, S., Stemmler, B., & López-Soriano, F.J. (2015). Cachexia and sarcopenia: mechanisms and potential targets for intervention. *Current Opinion in Pharmacology* 22, 100-106.
- Artioli, G.G., Sale, C., & Jones, R.L. (2018). Carnosine in health and disease. *European Journal of Sport Science*, 4, 1-10.
- Baugreet, S., Hamill, R.M., Kerry, J.P., & McCarthy, S.N. (2017). Mitigating nutrition and health deficiencies in older adults: A role for food innovation? *Journal of Food Science* 82, 848-855.
- Baum, J.I., Kim, I.Y., & Wolfe, R.R. (2016). Protein consumption and the elderly: What is the optimal level of intake? *Nutrients* 8(6), pii: E359.
- Binnie, M.A., Barlow, K., Johnson, V., & Harrison, C. (2014). Red meats: time for a paradigm shift in dietary advice. *Meat Science* 98, 445-451.
- Buffière, C., Gaudichon, C., Hafnaoui, N., Migné, C., Scislowky, V., Khodorova, N., Mosoni, L., Blot, A., Boirie, Y., Dardevet, D., Santé-Lhoutellier, V., & Rémond, D. (2017). In the elderly, meat protein assimilation from rare meat is lower than that from meat that is well done. *American Journal of Clinical Nutrition* 106(5), 1257-1266.
- Caldow, M.K., Ham, D.J., Godeassi, D.P., Chee, A., Lynch, G.S., & Koopman R. Glycine supplementation during calorie restriction accelerates fat loss and protects against further muscle loss in obese mice. *Clinical Nutrition* 35, 1118-1126.
- Cascella, M., Bimonte, S., Barbieri, A., Del Vecchio, V., Caliendo, D., Schiavone, V., Fusco, R., Granata, V., Arra, C., & Cuomo, A. (2018). Dissecting the mechanisms and molecules underlying the potential carcinogenicity of red and processed meat in colorectal cancer (CRC): an overview on the current state of knowledge. *Infectious Agents and Cancer* 13, 3.
- Clark, D.J., & Fielding, R.A. (2012). Neuromuscular contributions to age-related weakness. *Journals of Gerontology A Biological Sciences and Medical Sciences* 67(1), 41-47.

356 Cohen. S., Nathan, J.A., & Goldberg, A.L. (2015). Muscle wasting in disease: molecular
 357 mechanisms and promising therapies. *Nature Reviews in Drug Discovery* 14(1), 58-74.

358 Courtney-Martin, G., Ball, R.O., Pencharz, P.B., & Elango, R. (2016). Protein requirements
 359 during aging. *Nutrients* 8(8), pii: E492.

360 Cuthbertson, D., Smith, K., Babraj, J., Leese, G., Waddell, T., Atherton, P., Wackerhage, H.,
 361 Taylor, P.M., & Rennie, M.J. (2005). Anabolic signaling deficits underlie amino acid
 362 resistance of wasting, aging muscle. *FASEB Journal*, 19, 422-424.

363 Daly, R.M., O'Connell, S.L., Mundell, N.L., Grimes, C.A., Dunstan, D.W., & Nowson, C.A.
 364 (2014). Protein-enriched diet, with the use of lean red meat, combined with progressive
 365 resistance training enhances lean tissue mass and muscle strength and reduces circulating IL-
 366 6 concentrations in elderly women: a cluster randomized controlled trial. *American Journal*
 367 *of Clinical Nutrition* 99, 899-910.

368 Deshmukh-Taskar, P., Nicklas, T.A., Radcliffe, J.D., O'Neil, C.E., & Liu, Y. (2013). The
 369 relationship of breakfast skipping and type of breakfast consumed with overweight/obesity,
 370 abdominal obesity, other cardiometabolic risk factors and the metabolic syndrome in young
 371 adults. The National Health and Nutrition Examination Survey (NHANES): 1999-2006.
 372 *Public Health Nutrition* 16(11), 2073-2082.

373 Deutz, N.E., Bauer, J.M., Barazzoni, R., Biolo, G., Boirie, Y., Bosy-Westphal, A.,
 374 Cederholm, T., Cruz-Jentoft. A., Krznarić, Z., Nair, K.S., Singer, P., Teta, D., Tipton, K., &
 375 Calder, P.C. (2014). Protein intake and exercise for optimal muscle function with aging:
 376 recommendations from the ESPEN Expert Group. *Clinical Nutrition* 33, 929-936.

377 Evans, W.J. (2015). Sarcopenia should reflect the contribution of age-associated changes in
 378 skeletal muscle to risk of morbidity and mortality in elderly people. *Journal of the American*
 379 *Medical Directors Association* 16(7), 546-547.

380 Fazelzadeh, P., Hangelbroek, R.W., Tieland, M., de Groot, L.C., Verdijk, L.B., van Loon, L.
 381 J., Smilde, A., Alves, R.D., Vervoort, J., Muller, M., van Duynhoven, J.P., & Boekschoten,
 382 M.V. (2016). The muscle metabolome differs between healthy and frail older adults. *Journal*
 383 *of Proteome Research* 15, 499-509.

384 Fougere, B., Boulanger, E., Nourhashemi, F., Guyonnet, S., & Cesari, M. (2017). Chronic
 385 inflammation: Accelerator of biological aging. *Journals of Gerontology Series A Biological*
 386 *Sciences and Med Sciences* 72, 1218-1225.

387 Giesbertz, P., Padberg, I., Rein, D., Ecker, J., Hofle, A.S., Spanier, B., & Daniel, H. (2015).
 388 Metabolite profiling in plasma and tissues of *ob/ob* and *db/db* mice identifies novel markers
 389 of obesity and type 2 diabetes. *Diabetologia* 58, 2133-2143.

390 Ham, D.J., Caldow, M.K., Lynch, G.S., & Koopman, R. (2014a). Leucine as a treatment for
 391 muscle wasting: a critical review. *Clinical Nutrition* 33, 937-945.

392 Ham, D.J., Murphy, K.T., Chee, A., Lynch, G.S., & Koopman, R. (2014b). Glycine
 393 administration attenuates skeletal muscle wasting in a mouse model of cancer cachexia.
 394 *Clinical Nutrition* 33, 448-458.

395 Ham, D.J., Caldow, M.K., Lynch, G.S., & Koopman, R. (2014c). Arginine protects muscle
 396 cells from wasting in vitro in an mTORC1-dependent and NO-independent manner. *Amino*
 397 *Acids*. 46, 2643-2652.

398 Ham, D.J., Caldow, M.K., Chhen, V., Chee, A., Wang, X., Proud, C.G., Lynch, G.S., &
 399 Koopman, R. (2016). Glycine restores the anabolic response to leucine in a mouse model of
 400 acute inflammation. *American Journal of Physiology Endocrinology and Metabolism* 310,
 401 E970-E981.

402 Hepple, R.T., & Rice, C.L. (2016). Innervation and neuromuscular control in ageing skeletal
 403 muscle. *Journal of Physiology (London)* 594(8), 1965-1978.

404 Houtkooper, R.H., Argmann, C., Houten, S.M., Canto, C., Jeninga, E.H., Andreux, P.A.,
 405 Thomas, C., Doenlen, R., Schoonjans, K., & Auwerx, J. (2011). The metabolic footprint of
 406 aging in mice. *Scientific Reports* 1, 134.

407 Igase, M., Kohara, K., Okada, Y., Ochi, M., Igase, K., Inoue, N., Kutsuna, T., Miura, H., &
 408 Ohyagi, Y. (2018). A double-blind, placebo-controlled, randomised clinical study of the
 409 effect of pork collagen peptide supplementation on atherosclerosis in healthy older
 410 individuals. *Bioscience Biotechnology and Biochemistry*, Feb 15:1-3. doi:
 411 10.1080/09168451.2018.1434406.

412 Katsanos, C.S., Kobayashi, H., Sheffield-Moore, M., Aarsland, A. & Wolfe, R.R. (2005).
 413 Aging is associated with diminished accretion of muscle proteins after the ingestion of a
 414 small bolus of essential amino acids. *American Journal of Clinical Nutrition*, 82, 1065-1073.

415 Koopman, R., & van Loon, L.J. (2009). Aging, exercise and muscle protein metabolism.
 416 *Journal of Applied Physiology* 106, 2040-2048.

Koopman R, Crombach N, Gijsen AP, Walrand S, Fauquant J, Kies AK, Lemosquet S, Saris WH, Boirie Y, van Loon LJ (2009). Ingestion of a protein hydrolysate is accompanied by an accelerated in vivo digestion and absorption rate when compared with its intact protein. *American Journal of Clinical Nutrition* 90, 106-115.

Koopman R, Caldwor MK, Ham DJ, Lynch GS (2017). Glycine metabolism in skeletal muscle: implications for metabolic homeostasis. *Current Opinion in Clinical Nutrition and Metabolic Care* 20, 237-242.

Kouvaria, M., Tyrovolas, S., & Panagiotakos, D.B. (2016). Red meat consumption and healthy ageing: A review. *Maturitas* 84, 17-24.

Landi, F., Calvani, R., Tosato, M., Martone, A.M., Ortolani, E., Saveria, G., D'Angelo, E., Sisto, A., & Marzetti, E. (2016). Protein intake and muscle health in old age: From biological plausibility to clinical evidence. *Nutrients* 8(5) pii: E295.

Lang, C.H., & Frost, R.A. (2004). Differential effect of sepsis on ability of leucine and IGF-I to stimulate muscle translation initiation. *American Journal of Physiology Endocrinology and Metabolism* 287, E721-E730.

Lang, C.H., & Frost, R.A. (2005). Endotoxin disrupts the leucine-signaling pathway involving phosphorylation of mTOR, 4E-BP1, and S6K1 in skeletal muscle. *Journal of Cellular Physiology* 203, 144-155.

Lynch, G.S. (2011). *Sarcopenia – Age-Related Muscle Wasting and Weakness: Mechanisms and Treatments*, Springer Netherlands, 480 pp, ISBN: 978-90-481-9712-5.

McNeill, S.H. (2014). Inclusion of red meat in healthful dietary patterns. *Meat Science* 98, 452-460.

Millward, D.J., & Garnett, T. (2010). Food and the planet: nutritional dilemmas of greenhouse gas emission reductions through reduced intakes of meat and dairy foods. *Proceedings of the Nutrition Society* 69, 103-118.

Moon, A., Heywood, L., Rutherford, S., & Cobbold, C. (2013). Creatine supplementation: can it improve quality of life in the elderly without associated resistance training? *Current Aging Science* 6, 251-257.

Moore, D.R. (2014). Keeping older muscle “young” through dietary protein and physical activity. *Advances in Nutrition* 5, 599S-607S.

Morley, J.E., Anker, S.D., & von Haehling, S. (2014). Prevalence, incidence, and clinical impact of sarcopenia: facts, numbers, and epidemiology – update 2014. *Journal of Cachexia, Sarcopenia and Muscle* 5, 253-259.

Morris, M.S. & Jacques, P.F. (2013). Total protein, animal protein and physical activity in relation to muscle mass in middle-aged and older Americans. *British Journal of Nutrition* 109, 1294-1303.

Morton, R.W., Traylor, D., Weijs, P.J. M., & Phillips, S.M. (2018). Defining anabolic resistance: implications for delivery of clinical care nutrition. *Current Opinion in Critical Care* 24, 124-130.

Murton, A.J. (2015). Muscle protein turnover in the elderly and its potential contribution to the development of sarcopenia. *Proceedings of the Nutrition Society* 74(4), 387-396.

Ogawa, S., Yakabe, M. & Akishita, M. (2016). Age-related sarcopenia and its pathophysiological bases. *Inflammation and Regeneration* 36, 17.

Ortman, J., Velkoff, V., & Hogan, H. (2014). An aging nation: The older population in the United States. Retrieved from: <http://www.census.gov/prod/2014pubs/p25-1140.pdf>

Pennings, B., Boirie, Y., Senden, J.M., Gijsen, A.P., Kuipers, H., & van Loon, L.J. (2011). Whey protein stimulates postprandial muscle protein accretion more effectively than do casein and casein hydrolysate in older men. *American Journal of Clinical Nutrition* 93, 997-1005.

Pennings, B., Groen, B., de Lange, A., Gijsen, A.P., Zorenc, A.H., Senden, J.M., & van Loon, L.J. (2012). Amino acid absorption and subsequent muscle protein accretion following graded intakes of whey protein in elderly men. *American Journal of Physiology Endocrinology and Metabolism* 302, E992-E999.

Pennings, B, Groen, B.B., van Dijk, J.W., de Lange, A., Kiskini, A., Kuklinski, M., Senden, J.M., & van Loon, L.J. (2013). Minced beef is more rapidly digested and absorbed than beef steak, resulting in greater postprandial protein retention in older men. *American Journal of Clinical Nutrition* 98(1), 121-128.

Phillips, B.E., Williams, J.P., Greenhaff, P.L., Smith, K., & Atherton, P.J. (2017). Physiological adaptations to resistance exercise as a function of age. *JCI Insight* 2(17), pii: 95581.

477 Phillips, S.M. (2012). Nutrient-rich meat proteins in offsetting age-related muscle loss. *Meat*
478 *Science* 92, 174-178.

479 Piasecki, M., Ireland, A., Jones, D.A., & McPhee, J.S. (2016). Age-dependent motor unit
480 remodelling in human limb muscles. *Biogerontology* 17(3), 485-496.

481 Quintana Pacheco, D.A., Sookthai, D., Wittenbecher, C., Graf, M.E., Schübel, R., Johnson,
482 T., Katzke, V., Jakszyn, P., Kaaks, R., & Kühn, T. (2018). Red meat consumption and risk of
483 cardiovascular diseases – is increased iron load a possible link? *American Journal of Clinical*
484 *Nutrition* 2018 107, 113-119.

485 Rahman, M., Halade, G.V., El Jamali, A., & Fernandes G. (2009). Conjugated linoleic acid
486 (CLA) prevents age-associated skeletal muscle loss. *Biochemical and Biophysical Research*
487 *Communications* 383(4), 513-518.

488 Rennie, M.J. (2009). Anabolic resistance: the effects of aging, sexual dimorphism, and
489 immobilization on human muscle protein turnover. *Applied Physiology Nutrition and*
490 *Metabolism*, 34, 377-381.

491 Rohrmann, S., & Linseisen, J. (2016). Processed meat: the real villain? *Proceedings of the*
492 *Nutrition Society* 75(3), 233-241.

493 Rondanelli, M., Perna, S., Faliva, M.A., Peroni, G., Infantino, V., Pozzi, R. (2015). Novel
494 insights on intake of meat and prevention of sarcopenia: all reasons for an adequate
495 consumption. *Nutricion Hospitalaria* 32(5), 2136-2143.

496 Schaap, L.A., Pluijm, S.M., Deeg, D.J., Harris, T.B., Kritchevsky, S.B., Newman, A.B.,
497 Colbert, L.H., Pahor, M., Rubin, S.M., Tylavsky, F.A., Visser, M., & Health ABC Study
498 (2009). *Journals of Gerontology A Biological Sciences and Medical Sciences* 64(11), 1183-
499 1189.

500 Shaw, G., Lee-Barthel, A., Ross, M.L., Wang, B., and Baar, K. (2017). Vitamin C-enriched
501 gelatin supplementation before intermittent activity augments collagen synthesis. *American*
502 *Journal of Clinical Nutrition* 105, 136-143.

503 Short, K.R., Vittone, J.L., Bigelow, M.L., Proctor, D.N., & Nair, K.S. (2004). Age and
504 aerobic exercise training effects on whole body and muscle protein metabolism. *American*
505 *Journal of Physiology Endocrinology and Metabolism*, 286, E92-E101.

506 Sousa-Victor, P., & Muñoz-Cánoves, P. (2016). Regenerative decline of stem cells in
507 sarcopenia. *Molecular Aspects of Medicine* 50, 109-117.

508 Symons, T.B., Schutzler, S.E., Cocke, T.L., Chinkes, D.L., Wolfe, R.R., & Paddon-Jones, D.
 509 (2007). Aging does not impair the anabolic response to a protein-rich meal. *American*
 510 *Journal of Clinical Nutrition* 86(2),451-456.

511 Symons, T.B., Sheffield-Moore, M., Mamerow, M.M., Wolfe, R.R., & Paddon-Jones, D.
 512 (2011). The anabolic response to resistance exercise and a protein-rich meal is not diminished
 513 by age. *Journal of Nutrition, Health and Aging* 15(5), 376-381.

514 Tabung, F.K., Brown, L.S. & Fung, T.T. (2017). Dietary patterns and colorectal cancer risk:
 515 A review of 17 years of evidence (2000-2016). *Current Colorectal Cancer Reports*
 516 13(6),440-454.

517 Tieland, M., Borgonjen-Van den Berg, K.J., Van Loon, L.J., & de Groot, L.C. (2015).
 518 Dietary protein intake in Dutch elderly people: A focus on protein sources. *Nutrients*, 7(12),
 519 9697-9706.

520 Tintignac, L.A., Brenner, H.R., & Rüegg, M.A. (2015). Mechanisms regulating
 521 neuromuscular junction development and function and causes of muscle wasting.
 522 *Physiological Reviews* 95(3), 809-852.

523 Volpi, E., Sheffield-Moore, M., Rasmussen, B.B., & Wolfe, R.R. (2001). Basal muscle amino
 524 acid kinetics and protein synthesis in healthy young and older men. *Journal of the American*
 525 *Medical Association*, 286, 1206-1212.

526 Wall, B.T., Gorissen, S.H., Pennings, B., Koopman, R., Groen, B.B., Verdijk, L.B., & van
 527 Loon, L.J. (2015). Aging is accompanied by a blunted muscle protein synthetic response to
 528 protein ingestion. *PLoS ONE* 10, e0140903.

529 Welch, C., Hassan-Smith, Z.K., Greig, C.A., Lord, J.M. & Jackson, T.A. (2018). Acute
 530 sarcopenia secondary to hospitalisation – An emerging condition affecting older adults. *Aging*
 531 *and Disease* 9(1), 151-164.

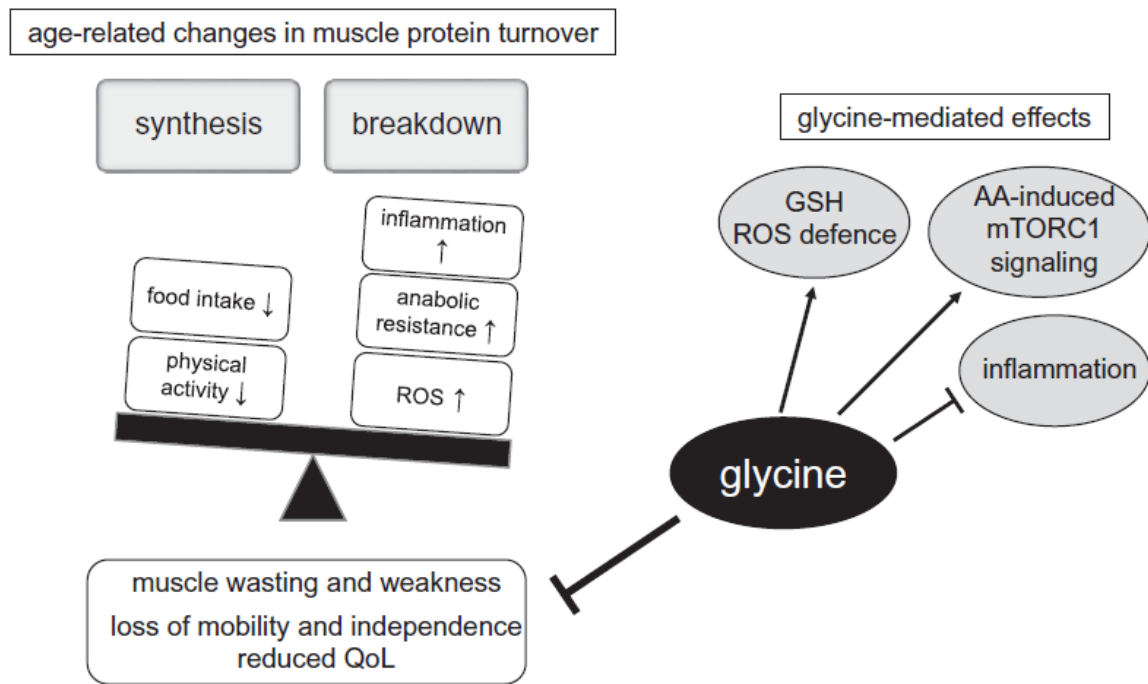
532 Whitham, M., Parker, B.L., Friedrichsen, M., Hingst, J.R., Hjorth, M., Hughes, W.E., Egan,
 533 C.L., Cron, L., Watt, K.I., Kuchel, R.P., Jayasooriah, N., Estevez, E., Petzold, T., Suter,
 534 C.M., Gregorevic, P., Kiens, B., Richter, E.A., James, D.E., Wojtaszewski, J.F.P., Febbraio,
 535 M.A. (2018). Extracellular vesicles provide a means for tissue crosstalk during exercise. *Cell*
 536 *Metabolism* 27(1), 237-251.

537 World Health Organization (2007). Protein and amino acid requirements in human nutrition

538 Report of a joint FAO/WHO/UNU expert consultation (WHO Technical Report Series 935).
539 ISBN: 92 4 120935 6.

540 Young, J.F., Therkildsen, M., Ekstrand, B., Che, B.N., Larsen, M.K., Oksbjerg, N., Stagsted,
541 J. (2013). Novel aspects of health promoting compounds in meat. *Meat Science* 95(4), 904-
542 911.

543



Legend for Figure 1.

Ageing is associated with an increase in inflammation and oxidative stress which impact on protein homeostasis; decreasing protein synthesis and increasing protein breakdown, leading to muscle wasting and weakness and contributing to the loss of independence and reduced quality of life (QoL). Glycine can attenuate muscle wasting by reducing inflammation, increasing defense mechanisms against reactive oxygen species, and increasing mTORC1 signaling. Glycine feeding or animal products with a high glycine content (such as gelatin), could represent simple and effective nutritional strategies for attenuating age-related muscle wasting and weakness.