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Genome-wide common and rare variant analysis provides novel insights into clozapine-associated neutropenia

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Conflict of Interest

L. Fredrik Jorskog has received research grant support from Teva/Auspex Pharmaceuticals and has served on a Data Safety and Monitoring Board for Janssen. David A Collier is a full-time employee and stockholder of Eli Lilly and Company Ltd.

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Abstract

The antipsychotic clozapine is uniquely effective in the management of schizophrenia, but its use is limited by its potential to induce agranulocytosis. The causes of this, and of its precursor neutropenia, are largely unknown although genetic factors play an important role. We sought risk alleles for clozapine-associated neutropenia in a sample of 66 cases and 5583 clozapine-treated controls, through a genome-wide association study (GWAS), imputed HLA alleles, exome array, and copy number variation analyses. We then combined associated variants in a meta-analysis with data from the Clozapine-Induced Agranulocytosis Consortium (up to 163 cases and 7970 controls). In the largest combined sample to date, we identified a novel association with rs149104283 (OR=4.32, $P=1.79 \times 10^{-8}$), intronic to transcripts of *SLCO1B3* and *SLCO1B7*, members of a family of hepatic transporter genes previously implicated in adverse drug reactions including simvastatin-induced myopathy and docetaxel-induced neutropenia. Exome array analysis identified gene-wide associations of uncommon non-synonymous variants within *UBAP2* and *STARD9*. We additionally provide independent replication of a previously identified variant in *HLA-DQB1* (OR=15.6, $P = 0.015$, positive predictive value = 35.1%). These results implicate biological pathways through which clozapine may act to cause this serious adverse effect.

Keywords

Clozapine; agranulocytosis; neutropenia; adverse effects; HLA-DQB1; SLCO1B3; SLCO1B7

Introduction

Clozapine is the only licensed medication for treatment-resistant schizophrenia (TRS), defined as a failure to respond to at least two antipsychotic trials of sufficient dose and duration. Although it is the only treatment with proven efficacy in this severely impaired group of patients, 1,2 it is substantially under-prescribed 3 due, at least in part, to the risk of haematological side effects of agranulocytosis and neutropenia (i.e., reductions of neutrophils to levels below 500/mm³ or 1500/mm³ respectively). The cumulative risk of agranulocytosis in those taking clozapine is 0.8% and for neutropenia is 2.9%. 4 If undetected, compromised immune function secondary to agranulocytosis can be fatal, as happened in a series of patients when the drug was introduced in the 1970s, leading to its widespread withdrawal. Evidence of its marked effectiveness over other antipsychotics led to Federal Drug Agency (USA) approval in 1989 with stipulations about the need for regular blood monitoring to aid early detection of blood abnormalities. The requirement for blood monitoring limits the acceptability of the drug to patients, and poses an obstacle to its use in clinical practice. 5

The aetiology of clozapine-induced blood disorders is currently unknown, though genetic causes contribute. The first genome-wide association study (GWAS) study conducted by the Clozapine-Induced Agranulocytosis Consortium (CIAC) has provided substantial evidence

for the role of *HLA-DQB1* and *HLA-B* in clozapine-associated neutropenia. 6 In this study we report analyses incorporating GWAS, HLA allele imputation, exome array and copy number variation to examine genetic associations with clozapine-associated neutropenia. Associated variants were combined in a joint meta-analysis with data from the CIAC study 6, giving the largest combined study sample of its kind to date.

Methods

Sample description

Study individuals were from CLOZUK (n=5493) and CardiffCOGS (Cognition in Schizophrenia, n=156) samples. All had clinical or research diagnoses of schizophrenia. 7 CLOZUK is comprised of individuals who were prescribed clozapine in the UK and have a clinical diagnosis of TRS. 7,8 The CLOZUK samples were acquired anonymously by the research team, in accordance with ethics permissions and the UK Human Tissue Act, in collaboration with Novartis, one of the UK suppliers of clozapine. Twelve months after sample acquisition, the research team were informed of those that had developed neutropenia whilst taking clozapine and where available, the recorded lowest neutrophil counts of these individuals were supplied. CardiffCOGS is a schizophrenia sample recruited from secondary mental health services in South Wales, UK; for detailed sample description see 8,9. As part of a comprehensive clinical interview, individuals were asked about lifetime clozapine use and occurrence of neutropenia. Clinical case notes were used to confirm neutropenia status and lowest recorded neutrophil levels were collected.

Clozapine-associated neutropenia cases (n=66) developed an absolute neutrophil count (ANC) $\leq 1500/\text{mm}^3$ during treatment with clozapine. Following the approach of recent studies, 6,10 we assessed cases with neutropenia because the success of the monitoring system and pre-emptive drug withdrawal in the UK has made agranulocytosis extremely rare. This neutrophil count threshold is used in the UK as a trigger to discontinue clozapine. Controls (n=5583) had received clozapine for a minimum of a year without developing an ANC $< 2000/\text{mm}^3$. Those who had a test result ($1500/\text{mm}^3 < \text{ANC} < 2000/\text{mm}^3$) were excluded from all analyses (n=20). No differences in age or sex were observed between clozapine-associated neutropenia cases and controls (Supplementary Table 1). All individuals were of European ancestry, as determined by self-report and principal component analysis of GWAS data.

Genotyping

Genotyping was performed at the Broad Institute, Cambridge, MA, USA. CardiffCOGS and part of the CLOZUK sample (40 cases and 3573 controls) were genotyped on Illumina HumanOmniExpressExome-8v1 and the remainder of the CLOZUK sample (26 cases and 2098 controls) were genotyped on both Illumina HumanOmniExpress-12v1 and Illumina HumanExome BeadChip.

Genome-wide association study

Quality control procedures and imputation were conducted using the Psychiatric Genomics Consortium (PGC) pipeline. 7 Imputation was performed using IMPUTE2 11 and a

reference panel from the full 1000 Genomes Project dataset (freeze date August 2012, see Supplementary Methods). Principal component estimation was conducted using EIGENSTRAT to exclude outliers and assess population stratification¹² (Supplementary Figures 1 and 2). We included genotyping array as well as the first three principal components as covariates to account for population structure. SNPs with allele frequencies that differed between genotyping arrays at $P < 1 \times 10^{-5}$ were excluded (Supplementary Figure 3). We selected common SNPs for analysis with high imputation quality (INFO > 0.8, MAF > 0.01 in cases and controls). Association analysis was performed using logistic regression in PLINK¹³ and SNPs functionally annotated using the Scripps genome advisor¹⁴. PLINK¹³ was used to identify index SNPs in relative linkage equilibrium. Further details of quality control procedures and statistical analyses are provided in Supplementary Methods.

HLA analysis

Classical *HLA* alleles and amino acid polymorphisms were imputed using SNP2HLA (version 1.02)¹⁵ using BEAGLE (version 3.0.4)¹⁶ from genotyped common variants using a reference dataset of 5225 individuals from Type 1 Diabetes Genetics Consortium (T1DGC). We used the same procedures for SNP selection, analysis and covariate selection (three PCAs derived from GWAS, plus genotyping array) as described for the GWAS analysis above. Due to complex and extended LD in the MHC, we did not identify index SNPs in relative linkage equilibrium.

We additionally genotyped a candidate SNP, *HLA-DQB1* 6672G>C¹⁷ (rs113332494), in 60 cases and 305 age and sex matched controls. This SNP was genotyped separately as it was not imputed with sufficient quality to be reported in the GWAS or HLA analyses, and was a strong candidate variant¹⁷. Genotyping was conducted at deCODE genetics using the Centaurus (Nanogen) platform¹⁸. Association with clozapine-associated neutropenia was tested using Fisher's exact test given low minor allele counts but to ensure there were no effect of population stratification, we also conducted a logistic regression including three PCAs derived from GWAS with 5×10^8 permutations to generate empirical p-values.

Exome array analysis

The Illumina exome array is designed to genotype uncommon-to-rare coding variants previously observed in whole exome sequencing studies. Exome array data were available for 57 cases and 4958 exposed controls. Full details of the quality control procedures are provided in an open access publication¹⁹ and Supplementary Methods. Principal component analysis was conducted using EIGENSTRAT¹² with 14,743 common exome array variants in relative linkage equilibrium (MAF ≥ 0.05 , $r^2 < 0.2$) to assess population structure and identify outliers (Supplementary Figure 4). Due to the relatively small case sample size in our study we did not apply a frequency filter to variants in this analysis. Single variant association was conducted using logistic regression in PLINK with the first 10 principal components included as covariates. Adaptive permutations (between 10 and 1×10^9) were used to generate empirical p-values in logistic regression analyses. PLINK¹³ was used to identify index SNPs in relative linkage equilibrium. To test for the effects of multiple functional variants in genes, we used SKAT-O²⁰ with 2×10^6 permutations, including the

first 10 principal components, for genes with at least two uncommon ($MAF < 0.05$), non-synonymous (missense, stop or splice) variants.

Copy Number Variation

The identification and quality control of copy number variation (CNV) for this sample has been previously described⁸ and is detailed in Supplementary Methods. CNVs were included if they had a frequency ≤ 0.01 , contained ≥ 10 probes, and were $\geq 100\text{kb}$ in length. Samples that passed both CNV and GWAS quality control (63 cases and 5456 controls) were used to test genes for enrichment of exon disrupting CNVs using a 2-sided Fisher's exact test. Deletions and duplications were analysed separately.

Secondary analysis of clozapine-associated neutropenia below $\leq 1000/\text{mm}^3$

We conducted secondary analyses on a subset of the more severely affected cases with $ANC \leq 1000/\text{mm}^3$ ($n=18$). A total of four of these cases had developed agranulocytosis ($ANC \leq 500/\text{mm}^3$). We assessed the association of single variants with clozapine-associated neutropenia below $\leq 1000/\text{mm}^3$ in GWAS ($N=18$), exome array ($N=16$) and HLA imputation ($N=18$) analyses. All analyses conducted were consistent with methods used for clozapine-associated neutropenia, described above.

Replication sample and meta-analysis

We obtained summary statistics for associated SNPs from a recently published study by the CIAC6. In this study Goldstein and colleagues conducted a comprehensive genetic association study in 163 clozapine-induced neutropenia cases (98 with $ANC < 500 \text{ mm}^{-3}$, 61 with $500 \leq ANC \leq 1000 \text{ mm}^{-3}$ and 4 with $ANC \leq 1500 \text{ mm}^{-3}$). The CIAC study included: GWAS (161 cases with clozapine-induced neutropenia, 249 clozapine exposed controls without neutropenia and 947 unexposed controls), exome array analysis (148 cases and up to 7970 unexposed controls), and classical *HLA* allele imputation (162 cases and 4319 unexposed controls). These datasets formed the replication samples for the current study and were combined with results for both (i) clozapine-associated neutropenia and (ii) neutropenia ≤ 1000 analyses. SNPs that were associated with clozapine-associated neutropenia at $P < 1 \times 10^{-4}$ from our GWAS, or $P < 0.05$ from our *HLA* variant analysis, were combined with the replication data in fixed-effects meta-analyses using PLINK to estimate a combined odds ratio weighted by the study's inverse standard error. If an index SNP was not present in the replication data, a proxy SNP in strong LD ($r^2 \geq 0.8$) was substituted and the s.e. weighted (s.e.w) to account for the lack of information: $\text{s.e.w} = \text{s.e.} / \sqrt{r^2}$.²¹ The variants that were associated with clozapine-associated neutropenia from exome array analyses with $P < 0.01$ were combined with the replication data in a p-value based method in METAL²², weighted by the square root of the total sample size. We used different p-value replication thresholds for the *HLA* and exome array to arrive at approximately the same number of variants.

Results

Figure 1 provides a summary of the study design and key results from each analysis.

Genome-wide association study

We performed a genome-wide association study of 7,559,010 genotyped and imputed common SNPs (QQ plot in Supplementary Figure 5, $\lambda_{GC} = 0.95$). Two SNPs were associated with clozapine-associated neutropenia at the genome-wide significant level of $P < 5 \times 10^{-8}$ (Supplementary Figure 6, Supplementary Data 1); rs80208670 on chromosome 13 (OR = 8.76, 95% CI: 4.21-18.25, $P = 6.51 \times 10^{-9}$) and rs77897117 on chromosome 1 (OR = 4.02, 95% CI: 2.46-6.57, $P = 4.60 \times 10^{-8}$). Our sample size had 80% power to detect an odds ratio (OR) > 4 for alleles with MAF > 0.10 at $P < 5 \times 10^{-8}$ (Supplementary Figure 7). The genome-wide significant SNPs from the discovery CLOZUK GWAS, rs80208670 and rs77897117, were not significantly associated in CIAC (OR = 1.69, $P = 0.27$ and OR = 0.67, $P = 0.28$, respectively).

In total, there were 266 independent ($r^2 < 0.1$) SNPs associated with clozapine-associated neutropenia at $P < 1 \times 10^{-4}$ and we sought replication of these SNPs in the CIAC sample (257 of these SNPs were available or had an appropriate proxy, Supplementary Data 1). Table 1 lists the 10 most strongly associated SNPs from the meta-analysis. One SNP on chromosome 12 surpassed the GWS threshold for association with clozapine-associated neutropenia (OR = 4.32, $P = 1.79 \times 10^{-8}$). rs149104283 is intronic to transcripts of *SLCO1B3* and *SLCO1B7* (solute carrier organic anion transporter family, member 1B3 and member 1B7) and was present in 7.37% of cases vs. 1.52% of controls in our sample and 4.20% of cases vs. 1.67% of controls in the CIAC sample. For consideration of how this may be translated to risk allele carrier status see Supplementary Results.

HLA analysis

No imputed classical *HLA* allele or amino acid polymorphism was associated with clozapine-associated neutropenia at the genome-wide significant level ($P < 5 \times 10^{-8}$) in either the discovery analysis (SNPs = 7,751) or combined meta-analysis (SNPs = 102) (Supplementary Data 2). It was not possible to impute the amino acid polymorphisms *HLA-DQB1* (126Q) and *HLA-B* (158T) implicated in the CIAC study⁶ with sufficient quality (INFO > 0.8) in the discovery sample.

We additionally genotyped a previously associated variant, *HLA-DQB1* 6672G>C 17 (rs113332494), in 60 cases and 305 age and sex matched controls as it was not imputed with sufficient quality. We found independent support for association of *HLA-DQB1* 6672G>C (OR = 15.6, 95% CI: 1.6 - 151.4, $P = 0.015$), replicating previous reports of association with clozapine-induced agranulocytosis.¹⁷ The association strengthened when considering only those with ANC below $\leq 1000/\text{mm}^3$ (OR = 38.1, 95% CI: 3.4 - 430.9, $P = 0.0079$). For the associated 'G' allele, we found three heterozygote carriers among 60 cases, and a single heterozygote in 305 controls. Lowest neutrophil counts were available for two of the three 'G' case carriers, both of whom had a neutrophil level $< 1000/\text{mm}^3$ (700 and 900). The association remained after adjusting for GWAS PCAs. For further consideration of possible population specific effects at this locus please refer to Supplementary Results.

Exome array analysis

In exome array analyses, no single variant exceeded a significance threshold of $P < 4.3 \times 10^{-7}$, corresponding to a Bonferroni correction for 115,000 variants tested in either the discovery or combined meta-analyses (Supplementary Data 3, QQ plot given in Supplementary Figure 8, $\lambda_{GC} = 1.11$). Table 2 lists the 10 most strongly associated exome variants from the meta-analysis. Of interest is rs1546308 ($P = 1.10 \times 10^{-6}$), a missense variant in *SLCO1B7* and intronic to *SLCO1B3*, that is 92kb from the SNP that emerged as the only genome-wide significant variant from GWAS meta-analysis. rs1546308 is predicted to be benign and was present in 15.8% of cases vs. 5.8% of controls in our sample and 9.3% of cases vs. 5.6% of controls in the CIAC sample.

Supplementary Table 3 displays the 10 most significantly associated genes (including the total allele count and number of variants contributing to each gene) based on the SKAT-O analysis of genes with at least two uncommon ($MAF < 0.05$), non-synonymous variants from the exome array. There was evidence of association for two genes that exceeded a threshold of $P < 2.5 \times 10^{-6}$ (corresponding to a Bonferroni correction of 20,000 genes tested 23) (QQ plot in Supplementary Figure 9); *UBAP2* on chromosome 9 ($P = 1.02 \times 10^{-7}$) and *STARD9* on chromosome 15 ($P = 2.85 \times 10^{-7}$). Due to differing analytical methods used by CIAC, it was not possible to combine our gene-based results in a joint analysis (see Supplementary Methods for detailed explanation).

Copy Number Variation

In a genome-wide analysis, no individual gene was significantly enriched for large, rare exonic CNVs that exceeded a significance threshold of $P < 2.5 \times 10^{-6}$ (Supplementary Data 4).

Associations with rs1546308 and rs149104283 are not independent

We investigated the independence of rs1546308 (missense variant in *SLCO1B7* and intronic in *SLCO1B3* from exome chip analysis) and rs149104283 (GWAS intronic variant in *SLCO1B3* and *SLCO1B7*) in samples with data available for both variants (55 cases and 4834 controls). The linkage disequilibrium (LD) between the two variants in the sample was $r^2 = 0.15$, $D' = 0.84$. In a conditional logistic regression, the strength of the association of rs1546308 with clozapine-associated neutropenia was attenuated from $OR = 3.00$ (95% CI: 1.735-5.189, $P = 8.40 \times 10^{-5}$) to $OR = 2.16$ (95% CI: 1.093-4.251, $P = 0.027$) after adjusting for rs149104283. Haplotype analysis did not strengthen the association signal. Thus, these two findings are not independent, and the associated region spans *SLCO1B1*, *SLCO1B3* and *SLCO1B7* (Figure 2, Supplementary Figure 14).

Secondary analysis of clozapine-associated neutropenia below $\leq 1000/\text{mm}^3$

Results of the GWAS for the more stringent neutropenia cut-off of $\leq 1000/\text{mm}^3$ are presented in Supplementary Results. The combined GWAS meta-analysis identified no loci exceeding genome-wide significance. The genome-wide significant SNP from the GWAS meta-analysis, rs149104283, had a minor allele frequency of 7.8% in cases with $ANC \leq 1000/\text{mm}^3$, and was associated at $OR = 7.6$, $P = 0.0027$ in the discovery sample. No variants

exceeded relevant significance thresholds in imputed HLA and exome array discovery or combined meta-analyses (see Supplementary Results).

Discussion

We have conducted a multifaceted genetic analysis of clozapine-associated neutropenia in the largest combined sample studied to date. Using GWAS, we identify a novel association implicating a family of organic anion transporters involved in drug metabolism which have been previously associated with adverse drug reactions. We also found evidence for effects of uncommon non-synonymous variants within *UBAP2* and *STARD9* and provide independent replication of a previously identified variant in *HLA-DQB1*.

The primary GWAS finding from the meta-analysis was a genome-wide significant association with neutropenia for rs149104283. The association effect size of this polymorphism is larger in the CLOZUK discovery sample (OR = 6.2) than in the CIAC replication dataset (OR = 2.95), as would be expected from winner's curse. It follows that the true effect size probably lies closer to the CIAC estimate, although this requires confirmation using independent data. rs149104283 is an intronic SNP for transcripts of both *SLCO1B3* and *SLCO1B7*; the associated region containing a third member of this organic anion transporter family, *SLCO1B1*. *SLCO1B7* encodes a putative protein (OAT1B7) that is poorly characterised, based on coding sequence prediction, and its functionality is unknown. *SLCO1B3* and *SLCO1B1* share sequence homology and encode liver-specific organic anion-transporter polypeptides (OATP1B3 and OATP1B1) that are multipass transmembrane proteins expressed exclusively in the basolateral membrane of hepatocytes. 24 They facilitate uptake of exogenous substances, including drugs, from the portal vein into hepatocytes, where the substance is subsequently modified either via metabolism with cytochrome (CYP) 450 enzymes or excreted. 25

Polymorphisms in *SLCO1B1* and *SLCO1B3* have been implicated in adverse reactions with other drugs. In 2008, a GWAS identified a missense variant rs4149056 in *SLCO1B1* that increased the risk of simvastatin-induced myopathy by increasing the area under the curve (AUC) for simvastatin, particularly in those taking high doses. 26 This prominent pharmacogenetic finding has been widely replicated and has led to recommendations for its use as a routine pre-emptive clinical test. 27 Particularly relevant to the current study are reports of an association between rs11045585, an intronic variant in *SLCO1B3*, and severe leukopenia/ neutropenia induced by docetaxel, a chemotherapeutic agent, 28 and that this may be secondary to alterations in the pharmacokinetics and bioavailability of the drug. 29–31 These polymorphisms were not in high LD ($r^2 < 0.1$ for both) with the index SNP in this study although rs11045585 was weakly associated with neutropenia in our discovery sample (OR=1.62, P=0.03).

Together, the findings suggest the hypothesis that genetic variants at *SLCO1B3* (and/or *SLCO1B1*) increase risk of clozapine-associated neutropenia through a pharmacokinetic mechanism. It is unclear whether clozapine plasma levels are associated with development of neutropenia. 32–34 One of the best-supported hypotheses to explain clozapine's association with agranulocytosis relates to the bioactivation of clozapine, or a stable

metabolite, to a chemically reactive nitrenium ion. 35 The propensity for nitrenium ions to cause apoptosis to neutrophils, or be toxic to neutrophil precursors, is dose dependent, lending support to the hypothesis that clozapine pharmacokinetics and bioavailability are related to its potential to cause neutropenia. 36,37

In analysis of exome chip data we found evidence of association with neutropenia for uncommon non-synonymous variants in *STARD9* and *UBAP2*. *STARD9* is a mitotic kinesin and *STARD9*-depleted cancer cells have abnormal cellular morphology and undergo apoptosis. 38 In addition, *STARD9*-depletion was found to synergise with the chemotherapeutic agent taxol, the use of which is dose-limited due to neutropenia. 38 The function of *UBAP2* is undetermined though it has an ubiquitin-associated domain and is widely expressed across tissues including bone marrow. The ubiquitination pathway has been shown to modulate the granulocyte colony-stimulating factor receptor 39,40, a critical regulator of neutrophil production. A recent study reported the association of a missense variant in the ubiquitin gene *USP43* with clozapine-associated neutropenia. 10

Our final finding adds to the growing evidence implicating *HLA-DQB1* in clozapine-associated neutropenia, supporting the recently published CIAC study. 6 There have been further reports implicating SNPs within *HLA-DQB1* 17,41,42 although these samples and those in CIAC are overlapping; thus we provide the first fully independent replication implicating this locus in clozapine-associated neutropenia/agranulocytosis. The *HLA-DQB1* variant alone has a positive predictive value of 35.1% (see Supplementary Table 4). Whilst this is promising, the majority of those that develop neutropenia or agranulocytosis whilst taking clozapine are not carriers of this risk allele, or indeed the other alleles we have identified in this study. The sensitivity for a test including rs149104283 (GWS intronic variant in *SLCO1B3* and *SLCO1B7*), rs1546308 (missense variant in *SLCO1B7*), and rs113332494 (*HLA-DQB1*) is 29.17%, the specificity 90.61%, the positive predictive value 9.94%, and the negative predictive value 97.30% (Supplementary Table 4). Although the variants identified in this study convey a substantially increased risk for clozapine-associated neutropenia, they are currently on their own unlikely to have clinical utility for pharmacogenetic testing 43, particularly as there is currently no alternative treatment for those with TRS.

An important consideration is that the majority of cases in our analyses had developed neutropenia rather than agranulocytosis. It is now very rare in the UK to develop agranulocytosis due to the success of the monitoring system and the fact that clozapine is stopped once neutropenia is detected; in fact only four cases met this threshold in our sample. Despite this we found that the major findings from our neutropenia analysis extended to the secondary analyses, which was restricted to those with an ANC $\leq 1000/\text{mm}^3$, indicating that the clozapine-associated neutropenia findings are likely to be applicable to those with severe neutropenia and agranulocytosis.

Our findings provide novel insights into putative biological processes underlying clozapine-associated neutropenia. Furthermore, we have indicated a potential link between the pharmacokinetics of clozapine and risk of neutropenia/agranulocytosis with potentially

important clinical implications. The development of such understanding should help widen the availability of clozapine with beneficial impact on those with TRS.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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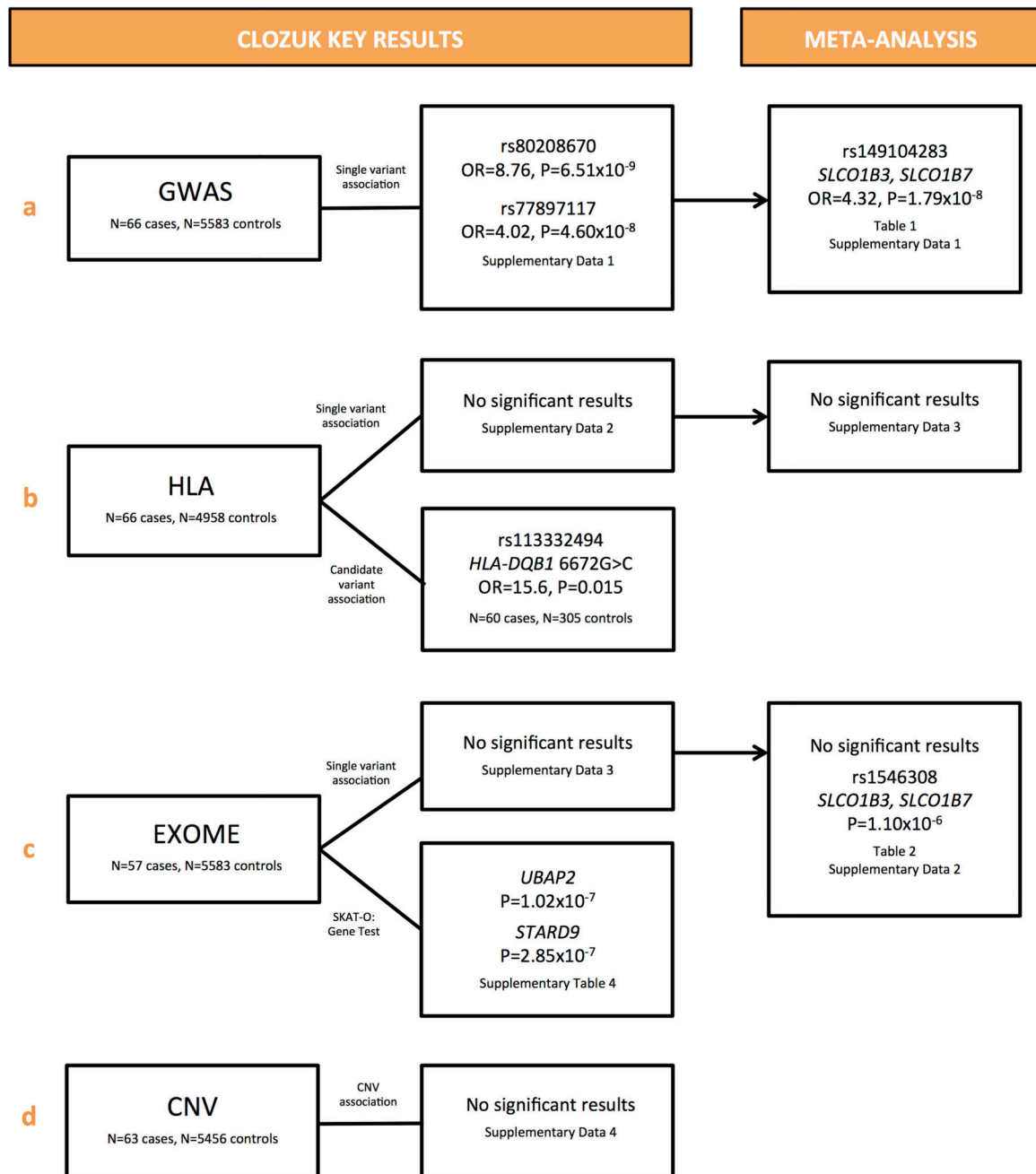


Figure 1. Study design and key results.

To investigate the association of genetic variants with clozapine-associated neutropenia, we conducted (a) a genome-wide association study (GWAS), (b) Human Leukocyte Antigen (HLA) allele imputation and genotyped a candidate variant of interest, HLA-DQB1 6672G>C/ rs113332494, (c) exome array single variant and gene-based analysis, and (d) copy number variation (CNV) analysis. We then took forward the associated variants from GWAS, HLA and exome array analyses to a combined meta-analysis with the Clozapine-Induced Agranulocytosis Consortium (CIAC) study.

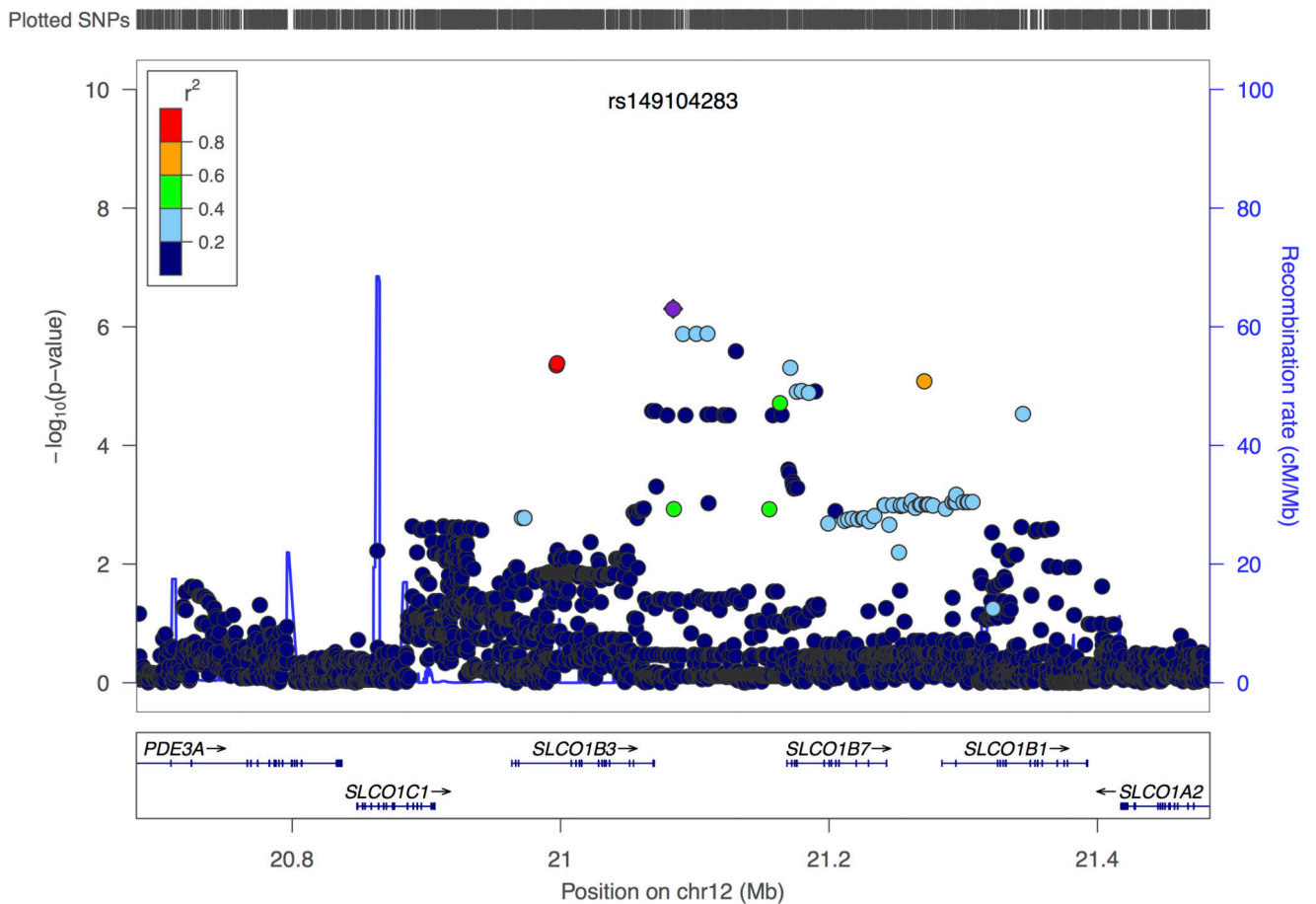


Figure 2. Association of 12p12.2 with clozapine-associated neutropenia.

LocusZoom plot of the region associated with clozapine-associated neutropenia on chromosome 12p12.2 in CLOZUK (discovery) sample. Genes within the region are shown in the lower panel, and the unbroken blue line indicated the recombination rate within the region. Each circle represents the P-value for one SNP in the discovery sample, with the top SNP rs149104283 shown in purple and the SNPs in the region coloured depending on their degree of correlation (r^2) with rs149104283 (as estimated by LocusZoom on the basis of CEU HapMap haplotypes).

Table 1

GWAS meta-analysis top 10 SNPs.

Results are ordered by meta-analysis P-value. Columns are: chromosome (CHR), variant ID (SNP), chromosomal position (Position), minor reference allele (A1), p-value, odds ratio (OR) and imputation quality (INFO) for CLOZUK (discovery) sample, CIAC (replication) sample, and meta-analysis, name of nearest gene and location to gene. Further details, including minor allele frequencies, are available in Supplementary Data 1.

CHR	SNP	Position	CLOZUK			CIAC			Meta-analysis			Nearest Gene	Location
			A1	P-value	OR	INFO	P-value	OR	INFO	P-value	OR		
12	rs149104283	21083862	T	4.98×10^{-7}	6.20	0.87	3.61×10^{-3}	2.95	0.87	1.79×10^{-8}	4.32	<i>SLCO1B3, SLCO1B7</i>	Intronic, intronic
13	rs80208670	84438088	C	6.51×10^{-9}	8.76	0.83	0.2728	1.69	0.83	1.56×10^{-7}	4.69	<i>SLITRK1</i>	13kb downstream
1	rs184597564	82236406	A	2.48×10^{-5}	4.03	0.89	4.99×10^{-3}	2.40	0.94	8.01×10^{-7}	3.06	<i>ADGRL2</i>	Intronic
7	rs78900159	76968378	A	6.94×10^{-6}	4.01	0.99	0.0247	1.97	0.98	2.02×10^{-6}	2.79	<i>GSAP</i>	Intronic
10	rs16916041	63146547	T	7.40×10^{-6}	2.57	1.02	0.0205	1.58	0.96	2.05×10^{-6}	1.98	<i>TMEM26</i>	20kb downstream
16	rs11649311	25226020	T	9.70×10^{-5}	2.07	0.89	2.62×10^{-3}	1.51	0.89	2.26×10^{-6}	1.68	<i>AQP8</i>	2kb upstream
17	rs117202297	53769035	T	8.48×10^{-6}	6.25	0.82	0.1503	2.66	0.66	5.27×10^{-6}	4.97	<i>TMEM100</i>	28kb downstream
17	rs80282661	13252073	T	1.99×10^{-6}	6.15	0.90	0.2632	1.85	0.89	5.49×10^{-6}	4.17	<i>HS3ST3A1</i>	147kb downstream
1	rs185053659	60704250	A	7.76×10^{-6}	5.80	0.86	0.1012	2.13	0.92	8.05×10^{-6}	3.80	<i>Clorf87</i>	165kb upstream
14	rs78074145	40404458	C	1.13×10^{-6}	4.05	0.96	0.2716	1.44	0.99	1.08×10^{-5}	2.60	<i>FXB033</i>	503kb upstream

Table 2

Exome array meta-analysis top 10 variants.

Results are ordered by meta-analysis P-value. Columns are: chromosome (CHR), variant as labelled by exome array (Variant), corresponding rs number for variant (Rs ID), chromosomal position, minor reference allele (A1), p-value for discovery (CLOZUK) sample, replication (CIAC) sample, and meta-analysis p-value, gene reference, location and function.

CHR	Variant	rs ID	Position	A1	CLOZUK P-value	CIAC P-value	Meta-analysis P-value	Gene	Location	Function
4	exm387558	rs201099591	7436363	A	2.48×10^{-3}	1.31×10^{-4}	1.10×10^{-6}	<i>PSAPL1, SORCS2</i>	Exonic, Intronic	Missense ¹
7	exm622984	rs17139320	63726370	G	3.96×10^{-3}	2.87×10^{-4}	3.70×10^{-6}	<i>ZNF679</i>	Exonic	Missense ¹
8	exm729894	rs201071539	145003862	A	2.19×10^{-3}	1.69×10^{-3}	1.19×10^{-5}	<i>PLEC</i>	Exonic	Missense ¹
19	exm1421170	rs2591594	9076728	A	4.66×10^{-3}	3.26×10^{-3}	4.52×10^{-5}	<i>MUC16</i>	Exonic	Missense ²
12	exm988839	rs1546308	21176135	C	1.25×10^{-4}	0.06786	9.13×10^{-5}	<i>SLCO1B7, SLCO1B3</i>	Exonic, Intronic	Missense ¹
1	exm23767	rs12073549	17720545	T	1.60×10^{-3}	0.01631	9.31×10^{-5}	<i>PADI6</i>	Exonic	Synonymous
15	exm1186658	rs117116488	89390513	T	2.76×10^{-4}	0.04477	9.73×10^{-5}	<i>ACAN</i>	Exonic	Missense ²
12	exm981950	rs79149293	8975873	G	5.66×10^{-5}	0.09674	1.01×10^{-4}	<i>A2ML1</i>	Exonic	Missense ³
12	exm995289	rs138912646	42711606	A	3.94×10^{-5}	0.1613	1.79×10^{-4}	<i>ZCRB1, PPHLN1</i>	Exonic, Intronic	Missense ¹
12	exm1051374	rs143584336	130921539	A	1.22×10^{-4}	0.1144	2.12×10^{-4}	<i>RIMBP2</i>	Exonic	Missense ¹

Predicted function of non-synonymous variants: ¹benign, ²possibly damaging, ³probably damaging. Further details, including minor allele frequencies, are available in Supplementary Data 3.